

低压低氧暴露对大鼠空间记忆及海马谷氨酸受体表达的影响*

付中伟 徐 玮 杜可军 戴 鹏 陈迎雷 黄冠鹏 蔡同建 骆文静 陈景元[△]

(第四军医大学军事预防医学院劳动与环境卫生教研室 陕西 西安 710032)

摘要 目的:观察低压低氧暴露对成年大鼠空间记忆及谷氨酸递质系统受体(AMPA,NMDA)的影响。方法:SD大鼠随机分成两组,对照组和低氧组(n=10),经过5天的Morris水迷宫训练,分别接受常压和低压低氧暴露7天,再通过Morris水迷宫观察暴露后的空间记忆,western blot检测GluR1 NMDA受体表达情况。结果:水迷宫结果显示低压低氧暴露后,平均逃脱潜伏期增长,平台搜索能力下降。Western blot结果显示磷酸化的GLUR1受体和NMDA受体水平升高。结论:低压低氧暴露可诱导大鼠的空间记忆损伤,其机制可能与谷氨酸递质系统紊乱造成的兴奋性中毒有关。

关键词 低压低氧;谷氨酸;空间记忆;GluR1;NMDA;兴奋性中毒

中图分类号 R821.5 文献标识码 A 文章编号:1673-6273(2012)15-2867-04

Effect of Hypobaric Hypoxia to The Spatial Memory and Glutamate Receptor*

FU Zhong-wei, XU Mei, DU Ke-jun, DAI Peng, CHEN Ying-lei,

HUANG Guan-peng, CAI Tong-jian, LUO Wen-jing, CHEN Jing-yuan[△]

(Department of Occupational and Environmental Health, Faculty of Military Preventive Medicine,
the Fourth Military Medical University, Shaanxi, Xi'an 710032, China)

ABSTRACT Objective: To investigate the effects of exposure to hypobaric hypoxia environment on spatial memory in adult rats and AMPA, NMDA receptors. **Methods:** Adult male SD rats were allocated randomized to the control and hypoxia groups. After five days of Morris Water Maze test training, two groups respectively exposed to normal atmospheric pressure or hypobaric hypoxia for 7days. The spatial memory was observed by morris water maze test. Western blot was used to detect the expression of GluR1 and NMDA. **Result:** In the Morris Water Maze test, the average escape latency increased and the ability to searching platform decreased significantly in the hypoxia group. Western blot results showed that the levels of p- GluR1 and NMDA rose. **Conclusion:** Exposing to hypobaric hypoxia leads to spatial memory damage in adult rats which may be related to the excitotoxicity resulted by glu transmitter system disorder.

Key words: Hypobaric hypoxia; Spatial memory; Glutamate; GluR1; NMDA; Excitotoxicity

Chinese Library Classification(CLC): R821.5 **Document code:** A

Article ID: 1673-6273(2012)15-2867-04

前言

高海拔低气压环境可对人体产生生理和心理上的影响,尤其是初到高原者,可能会产生剧烈的急性高原反应如头痛、恶心、失眠、眩晕,严重者发生急性高原病中高原脑水肿、高原肺水肿,其致死率很高^[1]。另外处于不同的海拔及不同的低压持续时间会对认知功能产生影响^[1,2]。人群研究和动物实验发现围产期低氧暴露会使智力发育滞后和发育不足^[3,4]。低压低氧暴露同样对成年大鼠的学习记忆功能产生不利影响^[5]。

在学习记忆产生的机制中,谷氨酸系统被认为有重要的意义。谷氨酸是神经系统内重要的神经递质。脑内的谷氨酸主要由谷氨酰胺被谷氨酰胺酶水解成Glu,发挥神经递质作用。离子型谷氨酸受体NMDA和代谢性谷氨酸受体AMPA是参与形成长时程增强(LTP)的主要受体^[6,7]。LTP被认为与学习记忆

有重要的联系^[8,9]。另外谷氨酸系统过度开放会引起神经兴奋性中毒,对脑功能产生不利影响^[10]。高原低压环境会对脑内神经递质系统产生影响^[11,12],本研究将探讨谷氨酸递质系统在低氧对学习记忆影响中的作用,研究这一问题将有助于了解低压低氧对脑影响的机制。

1 材料方法

1.1 实验动物和分组

选用20只SD雄性大鼠,体重200±25g,随机分为两组,对照组和低氧组(n=10)。动物饲养与室温,12小时照明~12小时无照明的动物房内,无限制食物与水饲养。

1.2 Morris水迷宫训练

水迷宫直径2m,深度0.5m,实验时装水约0.3m深,水温21~24℃,用白色素(二氧化钛)将水池染混。水迷宫空间人为分

* 基金项目 国家科技支撑计划(2009BA185B04)

作者简介 付中伟(1984-)男,硕士,研究方向为特种环境神经毒理,电话:029-84774866,E-mail:24007481@qq.com

△通讯作者 陈景元,Tel:029-84774401,E-mail:jy_chen@fmmu.edu.cn

(收稿日期 2012-01-18 接受日期 2012-02-15)

为4个象限(Q1-Q4,见图1.1),潜伏平台直径10 cm,置于Q1水面下约1.5 cm(如图1.1)。将大鼠置于潜伏平台(Q1)30 s后,按随机象限顺序置入水中60 s或大鼠找到潜伏平台为止,按此法每个象限训练2次。训练5天。

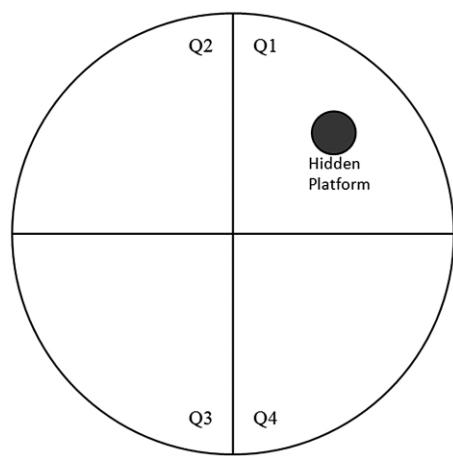


图1.1: Morris水迷宫示意图
Fig 1.1: Sketch of Morris Water Maze

1.3 低压低氧暴露

将低氧组大鼠置入低氧舱内,期间自由进食水,将气压控制在50 kPa水平(模拟5000~6000 m海拔),持续12小时后,移至动物房内(约100 kPa)。同时间对照组置于动物房(100 kPa)内,自由饮水。按此法暴露7天。

1.4 暴露后水迷宫检测

1.4.1 定位巡航测试 水迷宫内设定潜伏平台(Q1),按随机象限将大鼠置入水中60 s或大鼠找到潜伏平台为止,每个象限测试1次。观察暴露后2天(day13, day14)的水平。

1.4.2 平台搜索测试 暴露后第3天(day15),将大鼠从Q3置入无潜伏平台的水迷宫中,观察60 s内大鼠经过原平台次数及Q1搜索分布时间。

1.5 Western Blot

暴露后第4天(day16)将动物用1%戊巴比妥纳麻醉后,

断头处死,取脑组织分离海马,用碧云天公司提供的组织裂解液(含1%PMSF,SIGMA)按每1 mg加入10 μl裂解组织,充分裂解后4℃离心15 min(12000 r/min)后,取上清后将每组上清分别混合,进行蛋白定量,然后用裂解液配平浓度。最后加入2×SDS上样缓冲液后煮沸5 min、利用变性聚丙烯酰胺凝胶进行电泳分离,湿法将蛋白转移至PVDF膜,用5%TBST-脱脂牛奶进行室温封闭1 h。一抗4℃孵育过夜,用TBST洗涤3次,加入二抗室温孵育2 h。TBST洗涤3次,利用FluorChem FC2成像系统进行化学发光。

1.6 数据处理

用SPSS19.0软件进行统计分析,计量资料用均值±标准差($\bar{x} \pm s$)表示。两组间进行t检验比较,取 $P < 0.05$ 作为检验水准。

2 结果

2.1 低氧暴露前两组大鼠的水迷宫训练结果

如图2.1中day1~day5所示,通过5天训练,两组大鼠的逃脱潜伏期明显下降($P < 0.05$),但对照组与低氧组两组间无统计学差异。

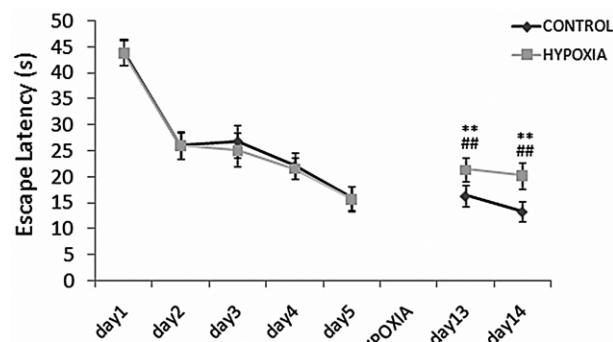


图2.1.大鼠低氧暴露前后水迷宫平均逃脱潜伏期(* $P < 0.05$ vs同日对照组数据, # $P < 0.05$ vs同组day5数据)

Fig. 2.1 Average escape Latency before/after exposure to hypoxia in MWM. (* $P < 0.05$ vs the control group on the same day, # $P < 0.05$ vs the same group on the day5)

表2.1 大鼠低氧暴露前后水迷宫平均逃脱潜伏期

Tabel 2.1 Average escape Latency before/after exposure to hypoxia in MWM

	Control	Hypoxia
Day1	44.05± 2.60	43.93± 2.47
Day2	26.01± 2.47	26.15± 2.72
Day3	26.84± 3.2	25.22± 3.25
Day4	22.12± 2.50	21.64± 2.07
Day5	16.02± 2.30	15.74± 2.42
Day13	16.36± 1.98	21.42± 2.19**
Day14	13.34± 1.88	20.27± 2.57**

Note: * $P < 0.05$ vs control.

2.2 暴露后水迷宫测试

通过7天低氧暴露,在暴露后1、2天(day13,14)的定位巡航实验中,低氧组大鼠的逃脱潜伏期较对照组有明显的升高

($P < 0.05$)(图2.1),同时低氧组暴露后较暴露前Day5逃脱潜伏期明显升高($P < 0.05$)(图2.1)。在暴露后第3天的平台搜索实验中,低氧组大鼠在经过平台次数(图2.2A)、Q1搜索时间分布较

对照组明显减少($P<0.05$)(图 2.2B),而对照组则无明显差异。两组间游泳速度无差别(图 2.2C)。这部分结果提示低氧暴露对大

鼠的空间记忆形成了损伤。

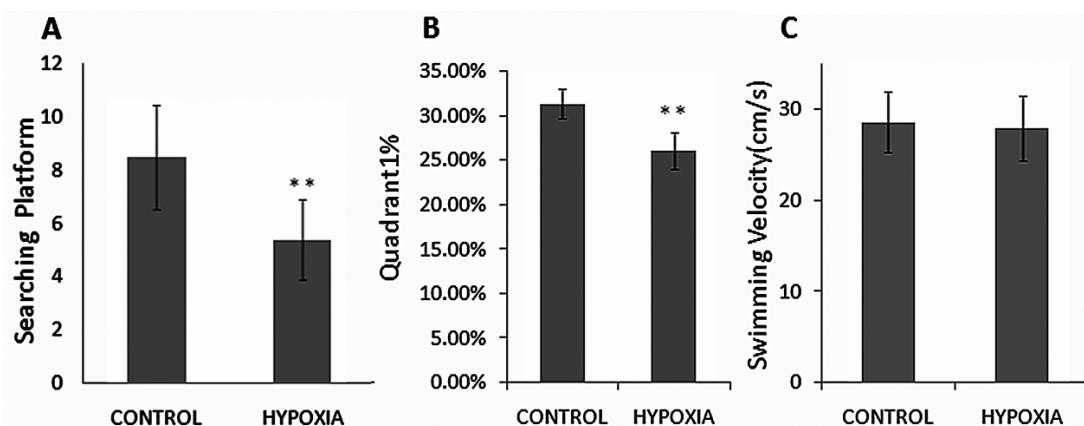


图 2.2 大鼠平台搜索实验各数据对比。A: 经过平台区域次数。B: Q1 分布百分比。C: 平均游泳速度对比。(* $P<0.05$ vs 对照组。)

Fig.2.2 Data in rats searching platform experiments. A: The Frequency of passing through the platform. B: The percentage of searching in the quadrant 1. C: The swimming velocity.(* $P<0.05$ vs the control group)

表 2.2 大鼠平台搜索实验各数据
Table 2.2 Data in rats searching platform experiments

	Control	Hypoxia
Frequency	8.5± 1.96	5.4± 1.51 **
Quadrant 1%	31.41± 1.72	26.09± 2.03 **
Swimming velocity (cm/s)	28.6± 3.37	27.9± 3.55

Note: * $P<0.05$ vs control group.

2.3 低氧暴露后大鼠海马组织内的 GluR1 磷酸化升高, NMDA 表达增高 受体中 GluR1 受体的 831,845 位点磷酸化均升高($P<0.05$), NMDA(NR1,NR2)表达量升高。

通过 7 天的低氧暴露, 检测发现大鼠海马组织内 AMPA

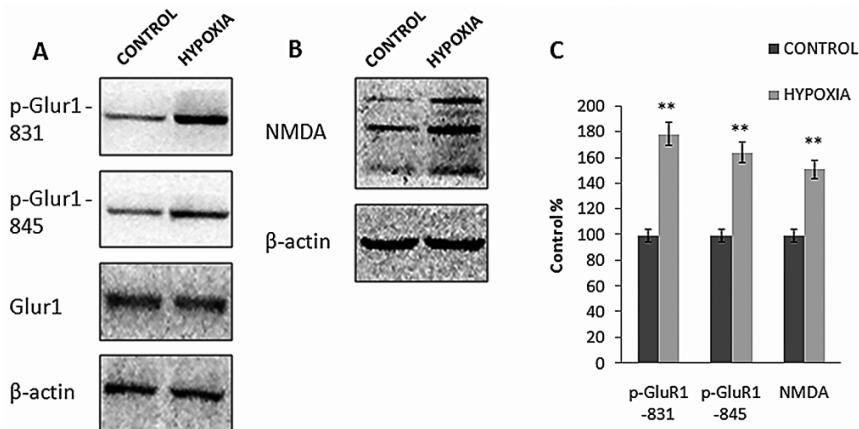


图 2.3 各组 GluR1 及 NMDA 受体表达情况。A: GluR1 831 和 845 位点磷酸化水平及总 GluR1 表达。B: NMDA 受体的表达情况。C: 灰度分析。(* $P<0.05$ vs 对照组。)

Fig. 2.3 GluR1 and NMDA expression. A p-GluR1(831,845)/GluR1 expression. B: NMDA expression. C: Quantification.(* $P<0.05$ vs the control group)

3 讨论

高原低压低氧对于认知功能有影响^[13], 低压低氧暴露的气压值和暴露时间与学习能力的降低呈高度依赖性^[13]。谷氨酸受体 AMPA 和 NMDA 在学习记忆形成的机制中十分重要^[6,7], 了解谷氨酸受体在低压环境中的变化有助于研究低压低氧对脑

的影响。

低压低氧暴露会引起脑内众多部位的神经元死亡, 如海马 CA1、CA3、齿状回、丘脑、皮质、纹状体^[14,15,16,17]。在行为学研究中, 19 天 7000 m 暴露水平会使新生大鼠的空间记忆产生严重的损伤^[18], 在成年大鼠中 2-6 小时的低压暴露就会使大鼠 Morris 水迷宫行为出现显著改变^[13], 但其中的机制还不明确。

低氧暴露会引起 NMDA 受体的开放呈先升高后降低的趋势^[19],长期暴露引起 NMDA 受体开放降低,从而影响 LTP 的形成,造成学习记忆的受损。然而在高原的初期,脑内处于低灌注水平,易造成兴奋性中毒^[20]。低氧造成神经细胞外液的 Glu 水平增高,抑制谷氨酸-胱氨酸转运系统,阻止其摄取胱氨酸用于合成谷胱甘肽^[21]。过度刺激谷氨酸受体,Glu 与 NMDA 持续结合,使得钙离子通道持续开放,导致钙离子内流,作用可持续 7 天左右,造成细胞死亡^[22]。另外兴奋性中毒造成的线粒体功能不全,大量产生氧自由基,可以使钙离子通道通透性进一步增加^[23]。线粒体形成的氧自由基又可以激活 NMDA 受体^[24],GluR 的过度活化同时可以诱导细胞凋亡^[25],钙离子可以激活核酸内切酶,蛋白激酶 C 等,诱导凋亡基因表达,这可能是其造成学习记忆损伤的一个原因。兴奋性毒性也可以诱导 Caspase-3 的表达^[26],同时激活凋亡相关蛋白 p53^[27]。

本研究中观察到低压低氧暴露后空间记忆的损伤,另外 GluR1 和 NMDA 受体较对照组显著增高,说明突触谷氨酸受体持续开放,提示神经元细胞兴奋性中毒。综上所述,在低氧暴露早期或急性期,由于谷氨酸递质系统紊乱,引起神经系统兴奋性中毒,造成神经元死亡,可能是低氧暴露诱导空间记忆损伤的机制。我们的结果有助于预防和保护高原低压低氧环境对于健康的影响。

参考文献(References)

- [1] Angerer P, Nowak D. Working in permanent hypoxia for fire protection-impact on health[J]. Int Arch Occup Environ Health, 2003, 76:87-102
- [2] Bartholomew CJ, Jenson W, Petros TV, Ferraro FR, Fire KM, Biberdorf D, et al. The effect of moderate levels of simulated altitude on sustained cognitive performance [J]. Int J Aviat Psychol, 1999, 9 (4):351-359
- [3] Askew EW. Work at high altitude and oxidative stress: antioxidant nutrients[J]. Toxicology, 2002, 180:107-119
- [4] Shukitt-Hale B, Kadar T, Marlowe BE, Stillman MJ, Galli RL, Levy A, et al. Morphological alterations in the hippocampus following hypobaric hypoxia[J]. Hum Exp Toxicol, 1996, 15(4):312-319
- [5] Cavallo G, Garavaglia P, Arragoni G, Tredici G. A quantitative three dimensional analysis of granule cell dendrites in the rat dentate gyrus [J]. J Comp Neurol, 1990, 302:206-219
- [6] Bliss TV, Lomo Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path[J]. J Physiol, 1973, 232:331,356
- [7] 刘巧凤,金清华,姜海英,等.习得性长时程增强中清醒大鼠海马齿状回区几种氨基酸的变化[J].中国临床康复,2006,10: 71-75
Liu Qiao-feng, Jin Qing-hua, Jiang Hai-ying, et al. Changes of amino acids in hippocampal dentate gyrus during learning-dependent long-term potentiation in free-moving conscious rats [J]. Chinese Journal of Clinical Rehabilitation, 2006, 10(38):71-75 (In Chinese)
- [8] 许琳,张均田.突触长时程增强形成机制的研究进展[J].生理学进展,2001,32:298-301
Xu Lin, Zhang Jun-tian. Advancement in Mechanisms of Long-Term Potentiation [J]. Progress in Physiological Sciences, 2001, 32 (4): 298-301 (In Chinese)
- [9] Molnár E. Long-term potentiation in cultured hippocampal neurons [J]. Semin Cell Dev Biol, 2011, 22(5):506-513
- [10] Szydlowska K, Tymianski M. Calcium, ischemia and excitotoxicity [J]. Cell Calcium, 2010 Feb;47(2):122-129
- [11] Tsai FS, Peng WH, Wang WH, Wu CR, Hsieh CC, Lin YT, et al. Effects of luteolin on learning acquisition in rats: involvement of the central cholinergic system[J]. Life Sci, 2007, 80(18):1692-1698
- [12] Row BW, Kheirandish L, Cheng Y, Rowell PP, Gozal D. Impaired spatial working memory and altered choline acetyltransferase (CHAT) immunoreactivity and nicotinic receptor binding in rats exposed to intermittent hypoxia during sleep [J]. Behav Brain Res, 2007, 177(2): 308-314
- [13] Shukitt-Hale B, Stillman MJ, Welch DL, et al. Hypobaric hypoxia impairs spatial memory in an elevation dependent fashion[J]. Behav Neuro Biol, 1996, 2(3):244-252
- [14] Freyaldenhoven TE, Ali SF, Schmued LC. Systemic administration of MPTP induced thalamic neuronal degeneration in mice [J]. Brain Res 1997, 759(1):9-17
- [15] Gibson GE, Duffy TE. Impaired synthesis of acetylcholine by mild hypoxic or nitrous oxide[J]. J Neurochem, 1981, 36:28-33
- [16] Naghdi N, Majlessi N, Bozorgmehr T. The effects of anisomycin (a protein synthesis inhibitor) on spatial learning and memory in CA1 region of rats hippocampus [J]. Behav Brain Res, 2003, 139 (1-2): 69-73
- [17] Smith P, Kesner RP, Chiba AA. Continuous recognition and spatial and non-spatial stimuli in hippocampal lesion rats [J]. Behav Neural Biol, 1993, 59:107-115
- [18] Simonova Z, Sterbova K, Brozek G, Komarek V, Sykova E. Postnatal hypo-baric hypoxia in rats impairs water maze learning and the morphology of neurones and macroglia in cortex and hippocampus[J]. Behav Brain Res, 2003, 141(2):195-205
- [19] Shukitt-Hale B, Stillman MJ, Welch DL, Levy A, Devine JA, Lieberman HR. Hypobaric hypoxia impairs spatial memory in an elevation-dependent fashion [J]. Behav Neural Biol, 1994, 62 (3): 244-252
- [20] 黄辉,阮怀珍,吴喜贵,等.低压低氧对大鼠海马神经元 NMDA 受体发育影响的研究[J].第三军医大学学报,2001,23(6): 655-657
Huang Hui, Ruan Huai-zhen, Wu Xi-gui, et al. Effect of hypobaric hypoxia on NMDA receptor of hippocampus neurons in developing rats [J]. Acta Academiae Medicinae Militaris Tertiae, 2001, 23 (6): 655-657 (In Chinese)
- [21] Pillis JW, Ren J, O'Regan MH. Transporter reversal as a mechanism of glutamate release from the ischemic rat cerebral cortex: studies with DL2-threo2beta2benzyl oxyaspartate[J]. BrainRes, 2000, 868:105-112
- [22] Valencia I, Mishra OP, Zubrow A, Fritz K, Katsetos CD, Delivoria-Papadopoulos M, Legido A. The role played by calcium in neuronal injury following neonatal hypoxia or convulsions [J]. Rev Neurol, 2006 Apr 10;42 Suppl 3:S11-15
- [23] LoPachin RM, Gaughan CL, Lehning EJ, Weber ML, Taylor CP. Effects of ion channel blockade on the distribution of Na, K, Ca and other elements in soxygen-glucose deprived CA1 hippocampal neurons[J]. Neuroscience, 2001, 103(4):971-983

(下转第 2825 页)

- cycle[J]. Biol Chem, 1956, 221(1):359-368
- [3] Sims CA, Wattanasirichaigoon S, Menconi MJ, et al. Ringer's ethyl pyruvate solution ameliorates ischemia/reperfusion-induced intestinal mucosal injury in rats[J]. Crit Care Med, 29(8):1513-1518
- [4] Cook VL, Holcombe SJ, Gandy JC, et al. Ethyl pyruvate decreases proinflammatory gene expression in lipopolysaccharide-stimulated equine monocytes [J]. Vet Immunol Immunopathol, 2011, 141(1-2): 92-99
- [5] Park SY, Yi EY, Jung M, et al. Ethyl pyruvate, an anti-inflammatory agent, inhibits tumor angiogenesis through inhibition of the NF- κ B signaling pathway[J]. Cancer Lett, 2011, 303(2): 150-154
- [6] Jang IS, Park MY, Shin IW, et al. Ethyl pyruvate has anti-inflammatory and delayed myocardial protective effects after regional ischemia/reperfusion injury[J]. Yonsei Med J, 2010, 51(6):838-844
- [7] Kao KK, Fink MP. The biochemical basis for the anti-inflammatory and cytoprotective actions of ethyl pyruvate and related compounds [J]. Biochem Pharmacol, 2010, 80(2): 151-159
- [8] Dong W, Cai B, Peña G, et al. Ethyl pyruvate prevents inflammatory responses and organ damage during resuscitation in porcine hemorrhage[J]. Shock, 2010, 34(2):205-213
- [9] Wang P, Gong G, Wei Z, et al. Ethyl pyruvate prevents intestinal inflammatory response and oxidative stress in a rat model of extrahepatic cholestasis[J]. J Surg Res, 2010, 160(2): 228-235
- [10] Davé SH, Tilstra JS, Matsuoka K, et al. Ethyl pyruvate decreases HMGB1 release and ameliorates murine colitis [J]. J Leukoc Biol, 2009, 86(3):633-643
- [11] Jang HJ, Kim YM, Tsoyi K, et al. Ethyl pyruvate induces HO-1 through p38 MAPK activation by depletion of glutathione in RAW 264.7 cells and improves survival in septic animals [J]. Antioxid Redox Signal, 2012 [Epub ahead of print]
- [12] Onur E, Akalin B, Memisoglu K, et al. Ethyl Pyruvate Improves Healing of Colonic Anastomosis in a Rat Model of Peritonitis[J]. Surg Innov, 2012 [Epub ahead of print]
- [13] Zhang J, Zhu JS, Zhou Z, et al. Inhibitory effects of ethyl pyruvate administration on human gastric cancer growth via regulation of the HMGB1-RAGE and Akt pathways in vitro and in vivo[J]. Oncol Rep, 2012, 27(5): 1511-1519
- [14] Hu X, Cui B, Zhou X, et al. Ethyl pyruvate reduces myocardial ischemia and reperfusion injury by inhibiting high mobility group box 1 protein in rats[J]. Mol Biol Rep, 2012, 39(1):227-231
- [15] Tian XX, Wu CX, Sun H, et al. Ethyl pyruvate inhibited HMGB1 expression induced by LPS in macrophages [J]. Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi, 2011, 27(12): 1304-1307, 1311
- [16] Yuan Y, Su Z, Pu Y, et al. Ethyl pyruvate promotes spinal cord repair through ameliorating the glial microenvironment [J]. Br J Pharmacol, 2011 [Epub ahead of print]
- [17] Chen HL, Bai H, Xi MM, et al. Ethyl pyruvate protects rats from phosgene-induced pulmonary edema by inhibiting cyclooxygenase2 and inducible nitric oxide synthase expression [J]. J Appl Toxicol, 2011 [Epub ahead of print]
- [18] Su X, Wang H, Zhao J, et al. Beneficial effects of ethyl pyruvate through inhibiting high-mobility group box 1 expression and TLR4/NF- κ B pathway after traumatic brain injury in the rat[J]. Mediators Inflamm, 2011, 2011: 807142
- [19] Huh SH, Chung YC, Piao Y, et al. Ethyl pyruvate rescues nigrostriatal dopaminergic neurons by regulating glial activation in a mouse model of Parkinson's disease[J]. J Immunol, 2011, 187(2): 960
- [20] Kung CW, Lee YM, Cheng PY, et al. Ethyl pyruvate reduces acute lung injury via regulation of iNOS and HO⁻¹ expression in endotoxemic rats[J]. J Surg Res, 2011, 167(2):323-331
- [21] Di Paola R, Mazzon E, Galuppo M, et al. Ethyl pyruvate therapy attenuates experimental severe arthritis caused by type II collagen (CII) in the mouse (CIA)[J]. Int J Immunopathol Pharmacol, 2010, 23(4): 1087-1098
- [22] Moro N, Sutton RL. Beneficial effects of sodium or ethyl pyruvate after traumatic brain injury in the rat [J]. Exp Neurol, 2010, 225(2): 391
- [23] Yu DH, Noh DH, Song RH, et al. Ethyl pyruvate downregulates tumor necrosis factor alpha and interleukin (IL)-6 and upregulates IL-10 in lipopolysaccharide-stimulated canine peripheral blood mononuclear cells[J]. J Vet Med Sci, 2010, 72(10): 1379-1381
- [24] Mizutani A, Maeda N, Toku S, et al. Inhibition by ethyl pyruvate of the nuclear translocation of nuclear factor- κ B in cultured lung epithelial cells[J]. Pulm Pharmacol Ther, 2010, 23(4): 308-315
- [25] Shen H, Hu X, Liu C, et al. Ethyl pyruvate protects against hypoxic-ischemic brain injury via anti-cell death and anti-inflammatory mechanisms[J]. Neurobiol Dis, 2010, 37(3): 711-722
- [26] Shang GH, Lin DJ, Xiao W, et al. Ethyl pyruvate reduces mortality in an endotoxin-induced severe acute lung injury mouse model [J]. Respir Res, 2009, 10: 91
- [27] Varma SD, Devamanoharan PS, Ali AH. Devamanoharan PS and Ali AH. Prevention of intra-cellular oxidative stress to lens by pyruvate and its ester[J]. Free Rad Res, 1998, 28: 131-135

(上接第 2870 页)

- [24] Dugan LL, Sensi SL, Canzoniero LM. Mitochondrial production of reactive oxygen species in cortical neurons following exposure to N2 methyl 2D2aspartate[J]. J Neurosci, 1995, 15(10):6377-6388
- [25] Raghupathi R, Graham D I. Mc Intosh TKApop tosis after traumatic brain injury[J]. J Neurotrauma, 2000, 17(10):927-938
- [26] Du Y, Dodel RC, Bales KR. Involvement of a caspase23like cysteine protease in 1-methyl-4-phenylpyridinium-mediated apoptosis of cultured cerebellar granule neurons [J]. J Neurochem, 1997, 69(4): 1382-1388
- [27] Meek DW. Multisite phosphorylation and the integration of stress signals at p53[J]. Cell Signal, 1998, 10(3):159-166