

doi: 10.13241/j.cnki.pmb.2015.11.007

苦参碱联合顺铂对小鼠宫颈癌 U14 移植瘤抑制作用的影响

李丹¹ 余爱云² 余亚娟³ 谢珊艳⁴ 刘怡安¹

(1 武汉科技大学附属天佑医院妇产科 湖北 武汉 430064;2 湖北科技学院妇产科 湖北 武汉 437100;3 江苏省无锡市人民医院
图室 江苏 无锡 214000;4 浙江省嘉兴市妇幼保健院病理科 浙江 嘉兴 314000)

摘要 目的:探讨苦参碱联合顺铂对小鼠宫颈癌 U14 移植瘤的抑制作用。**方法:**制备宫颈癌移植瘤小鼠模型,随机分为苦参碱高、中、低剂量组,顺铂联合苦参碱高、中、低剂量组,顺铂组及对照组 8 组,给予不同治疗后检测计算肿瘤抑瘤率,胸腺指数,脾脏指数,肌酐及尿素氮水平。**结果:**①相比对照组,其余各组瘤重均有减少,其中顺铂组、苦参碱高剂量组及联合用药各组瘤重减少明显,差异有统计学意义($P < 0.01$);②相比对照组,顺铂组胸腺指数及脾脏指数显著下降($P < 0.01$),相比对照组及顺铂组,苦参碱组与联合顺铂组胸腺指数及脾脏指数均增高,差异有显著性意义($P < 0.01$);③联合高剂量组肌酐、尿素氮值相比顺铂组显著降低($P < 0.01$),联合中剂量组相比顺铂组减少($P < 0.05$),低剂量组下降不明显($P > 0.05$)。**结论:**一定浓度的苦参碱对小鼠宫颈癌 U14 移植瘤有抑制作用,同顺铂联合应用可增强顺铂的抗肿瘤效果,并减轻后者的毒副作用,对机体免疫力提升亦有促进。

关键词:苦参碱;顺铂;宫颈癌;U14

中图分类号:Q95-3;R737.33 文献标识码:A 文章编号:1673-6273(2015)11-2027-04

Inhibitory Effect of Matrine Combined Cisplatin on Murine Cervical Cancer of U14 Xenograft

LI Dan¹, YU Ai-yun², YU Ya-juan³, XIE Shan-yan⁴, LIU Yi-an¹

(1 Department of Obstetrics and Gynecology, Tianyou Hospital Affiliated to Wuhan University of Science and Technology, Wuhan, Hubei, 430064, China; 2 Department of Obstetrics and Gynecology, Hubei Institute of Technology, Wuhan, Hubei, 437100, China;

3 Examination room, Wuxi people's Hospital of Jiangsu province, Wuxi, Jiangsu, 214000, China; 4 Department of Pathology, Jiaxing Maternal and Child Health-Care Center of Zhejiang Province, Jiaxing, Zhejiang, 314000, China)

ABSTRACT Objective: To explore the inhibitory effect of Matrine combined cisplatin on murine cervical cancer by U14 xenograft.

Methods: Cervical cancer transplantation tumor model in mice was perpetrated, all mice were randomly divided into 8 groups: matrine groups(high, medium and low dose group), cisplatin plus bitter and alkali groups(high, medium and low dose group), cisplatin group and control group. After treatment, tumor inhibition rate, thymus index, spleen index, creatinine and urea nitrogen levels were calculated.

Results: (1) Compared with the control group, tumor weight of other groups were reduced, in the cisplatin group, the matrine high-dose group and combination group, tumor weight decreased obviously, the difference was statistically significant($P < 0.01$). (2)Compared with control group, the thymus index and spleen index of cisplatin group decreased significantly ($P < 0.01$), compared with the control group and cisplatin group, thymus index and spleen index of matrine and combined cisplatin group increased significantly, with significant differences($P < 0.01$). (3) Compared with cisplatin group, creatinine and urea nitrogen values of high dose combination group decreased significantly ($P < 0.01$), values for medium dose combination group also decreased ($P < 0.05$), but no significant decrease was observed in low dose group ($P > 0.05$). **Conclusion:** A certain concentration of bitter and alkali has inhibitory effect on murine cervical cancer of U14 xenograft. When combined in treatment, it can enhance the anti-tumor effect of cisplatin, and reduce the side effects of the cisplatin, the immunity can be also promoted as well.

Key words: Matrine; Cisplatin; Cervical cancer; U14

Chinese Library Classification(CLC): Q95-3; R737.33 Document code: A

Article ID: 1673-6273(2015)11-2027-04

前言

随着现代医学及传统医学的不断进步和发展,中药在肿瘤

作者简介:李丹(1977-),女,硕士,主治医师,从事宫颈癌防治方面的研究,E-mail:lidan2236@126.com

(收稿日期:2014-11-15 接受日期:2014-12-05)

治疗中的作用受到越来越多的重视,利用或联合中药治疗肿瘤成为近年来抗癌治疗的新途径,苦参碱是具有广谱抗癌作用的中药提取物质,为探讨其对人类宫颈癌肿瘤细胞的影响,我们通过建立小鼠宫颈癌 U14 移植瘤模型进行了研究,现将结果报道如下:

1 材料和方法

1.1 材料

1.1.1 实验动物 清洁级雌性昆明种小鼠(购自兰州大学基础医学院实验动物中心,合格证号为 SCXX (甘)2009-0004)80只,体重 20 ± 3 g,6~8周龄。瘤细胞株:小鼠宫颈癌细胞株(U14),购于中国医学科学院北京药物研究所。

1.1.2 实验药物 注射用苦参碱(商品名甘缘,江苏康缘药业股份有限公司生产,规格0.15 g/支,批准文号国药准字H20041496);注射用顺铂冻干型(齐鲁制药有限公司,规格10 mg/支,批准文号国药准字H20023460);灭菌注射用水(华北制药股份有限公司,规格5 mL/支,批准文号国药准字H13023195)。

1.1.3 实验试剂 于实验室自行配制DMEM高糖型培养液,0.1 mol/L PBS液(pH=7.4),0.25%及0.125%的胰蛋白酶,谷氨酰胺溶液,双抗(青霉素、链霉素),0.4%台盼蓝液,细胞冻存液;因小鼠平均给药体积为10 mL/kg,故分别配制浓度为75 mg/kg、50 mg/kg及35 mg/kg的苦参碱药液与2 mg/kg的顺铂药液。

1.2 方法

1.2.1 小鼠宫颈癌细胞株(U14)培养 将购入的U14细胞株接种并分装至DMEM高糖型培养液中,待其生长并铺满瓶底约80%时进行细胞传代,并在细胞培养过程中对其进行冻存和复苏,在冻存时将所有细胞制成细胞悬液,保存于-80℃冰箱中,复苏时应避免冰晶产生致细胞破碎。

1.2.2 小鼠宫颈癌细胞株(U14)腹腔传代 将处于对数期生长的U14细胞制成瘤细胞悬液,于雌性昆明小鼠腹腔内注射0.2 mL细胞悬液并传2~3代作为种鼠,抽取腹水以保种或荷瘤。

1.2.3 制备宫颈癌移植瘤小鼠模型 对所有小鼠进行称重并标记,随机分为8组,每组10只,在无菌条件下将从种鼠中抽取的腹水用生理盐水稀释,调整细胞浓度为 5×10^6 个/mL,制成瘤细胞悬液。在每只小鼠右前肢腋下靠腋中线处上方2 cm位置接种0.2 mL约 1×10^6 个活细胞,制成局灶性移植瘤模型。

1.3 治疗及处理

1.3.1 给药 8组分别为苦参碱高、中、低剂量组,顺铂联合苦参碱高、中、低剂量组,顺铂组及对照组。对照组给予灭菌注射用水,苦参碱高、中、低剂量组分别予75 mg/kg·d,50 mg/kg·d,35 mg/kg·d的剂量给药,顺铂组予顺铂2 mg/kg·d剂量给药,顺铂联合苦参碱高、中、低剂量组在苦参碱各剂量基础上联合顺铂2 mg/kg·d。各组小鼠均于瘤体周围局部注射给药,剂量均为0.2 mL,连续治疗10 d,治疗期间常规饲养,观察小鼠活动、进食及瘤体变化。

1.3.2 称重 在治疗11 d时对小鼠称重,计算体重均值,然后将小鼠脱臼处死,剥取移植瘤称重,计算肿瘤抑瘤率,胸腺指数,脾指数;肿瘤抑瘤率=(对照组平均瘤重-实验组平均瘤重)/对照组平均瘤重×100%;胸腺指数=(胸腺重量mg/体重g);脾指数=(脾重量mg/体重g)。

1.3.3 测定肌酐、尿素氮水平 摘取小鼠眼球采血后置于室温下2 h,离心后取血清,采用全自动生化分析仪测定尿素氮、肌酐水平。

1.4 统计方法

采用SPSS13.0统计软件进行数据分析,计量资料采用均数±标准差($\bar{x}\pm s$)表示,计量资料比较采用单因素方差分析,计数资料比较采用 χ^2 检验。以P<0.05认为差异有统计学意义。

2 结果

2.1 不同治疗对U14荷瘤小鼠移植瘤大小、瘤重及抑瘤率的影响

相比对照组,其余各组瘤重均有减少,其中顺铂组、苦参碱高剂量组及联合用药各组瘤重减少明显,差异有统计学意义(P<0.01),顺铂联合苦参碱高剂量组抑瘤率相比顺铂组较高,联合组各组抑瘤率相比苦参碱组均高。按照《现代肿瘤治疗药物学》抑瘤率>30%即为阳性的标准,表明苦参碱高、中剂量组对U14移植瘤有不同程度抑制,而联合顺铂后抗肿瘤效果更优于苦参碱组。见表1。

表1 不同治疗对U14荷瘤小鼠移植瘤大小、瘤重及抑瘤率的影响($\bar{x}\pm s$)

Table 1 Effect of different treatment on tumor size, weight and inhibition rate of murine U14 tumor xenografts($\bar{x}\pm s$)

组别 Groups	初始体重(g) Initial weight(g)	终体重(g) Final weight(g)	瘤重(g) Tumor weight(g)	抑瘤率(%) Inhibition rate(%)
苦参碱高剂量组 Matrine high dose group	19.32± 0.88	26.51± 1.68	1.14± 0.52 [#]	38.71
苦参碱中剂量组 Matrine medium dose group	19.34± 0.87	26.54± 1.72	1.25± 0.68 [*]	33.12
苦参碱低剂量组 Matrine low dose group	19.33± 0.79	26.77± 1.69	1.58± 0.67	15.48
联合高剂量组 Combination with high dose group	19.65± 0.86	26.49± 1.66	0.77± 0.28 [#]	58.64
联合中剂量组 Combination with medium dose group	19.57± 0.87	26.48± 1.71	0.95± 0.39 [#]	49.62
联合低剂量组 Combination with low dose group	19.51± 0.83	26.49± 1.73	0.81± 0.33 [#]	42.13
顺铂组 Cisplatin group	19.46± 0.82	26.52± 1.69	1.87± 0.82	58.11
对照组 Control group	19.48± 0.85	26.64± 1.73	-	-

注:与对照组相比,*P<0.05,#P<0.01。

Note: compared with control group,*P<0.05,#P<0.01

2.2 苦参碱组与联合顺铂组对 U14 小鼠脏器指数的影响

相比对照组, 顺铂组胸腺指数及脾脏指数显著下降 ($P <$

0.01), 相比对照组及顺铂组, 苦参碱组与联合顺铂组胸腺指数及脾脏指数均增高, 差异有显著性意义 ($P < 0.01$), 见表 2。

表 2 苦参碱组与联合顺铂组对 U14 小鼠脏器指数的影响 ($\bar{x} \pm s$)

Table 2 Effect of Matrine group and cisplatin- combined group on U14 mice Viscera index ($\bar{x} \pm s$)

组别 Groups	剂量[mg/(kg·d)] Dose[mg/(kg·d)]		胸腺指数(mg/g)	脾脏指数(mg/g)
	苦参碱 Matrine	顺铂 Cisplatin	Thymus index(mg/g)	Spleen index(mg/g)
苦参碱高剂量组 Matrine high dose group	75	-	4.11± 0.37 ^{#△}	13.01± 2.77 ^{#△}
苦参碱中剂量组 Matrine medium dose group	50	-	3.31± 0.24 ^{#△}	12.23± 1.76 ^{#△}
苦参碱低剂量组 Matrine low dose group	35	-	2.94± 0.82 ^{*△}	11.02± 1.33 ^{*△}
联合高剂量组 Combination with high dose group	75	2	3.97± 0.13 ^{#△}	12.94± 0.23 ^{#△}
联合中剂量组 Combination with medium dose group	50	2	3.06± 0.55 ^{#△}	12.11± 2.74 ^{#△}
联合低剂量组 Combination with low dose group	35	2	2.31± 0.72	10.84± 0.86
顺铂组 Cisplatin group	-	-	2.12± 0.39	10.49± 1.66
对照组 Control group	-	2	0.92± 0.31 [#]	8.24± 1.19 [#]

注:与对照组相比,* $P < 0.05$,# $P < 0.01$,与顺铂组相比,^{*} $P < 0.05$,[△] $P < 0.01$ 。

Note: compared with control group, * $P < 0.05$, # $P < 0.01$, compared with Cisplatin group, ^{*} $P < 0.05$, [△] $P < 0.01$.

2.3 顺铂组与联合顺铂组对 U14 小鼠肌酐、尿素氮水平比较

联合高剂量组肌酐、尿素氮值相比顺铂组显著降低 ($P <$

0.01), 联合中剂量组相比顺铂组减少 ($P < 0.05$), 低剂量组下降不明显 ($P > 0.05$)。具体见表 3。

表 3 顺铂组与联合顺铂组 U14 小鼠肌酐、尿素氮水平比较 ($\bar{x} \pm s$)

Table 3 Comparison of creatinine and urea nitrogen levels in U14 mice among cisplatin group and cisplatin-combined group ($\bar{x} \pm s$)

组别 Groups	剂量[mg/(kg·d)] Dose[mg/(kg·d)]		肌酐(μmol/L)	尿素氮(μmol/L)
	苦参碱 Matrine	顺铂 Cisplatin	Creatinine (μmol/L)	Urea nitrogen levels (μmol/L)
联合高剂量组 High dose combination group	75	2	59.11± 3.68 [#]	8.71± 1.12 [#]
联合中剂量组 Medium dose combination group	50	2	61.09± 3.76 [#]	9.92± 0.73 [*]
联合低剂量组 Low dose combination group	35	2	64.55± 3.67 [*]	10.83± 0.81
顺铂组 Cisplatin group	-	2	72.14± 3.81	12.24± 2.55

注:与顺铂组相比,* $P < 0.05$,# $P < 0.01$ 。

Note: compared with cisplatin group, * $P < 0.05$, # $P < 0.01$.

3 讨论

既往实验研究已证实苦参碱具有广谱抑制肿瘤细胞的作用, 其机体毒性低, 可通过升高白细胞等提高机体免疫力, 从而有利于患者对抗肿瘤^[1-5]。本组研究中将 U14 荷瘤小鼠为模型, 探讨苦参碱及苦参碱联合顺铂对 U14 移植瘤的抑制作用。结果表明, 苦参碱高剂量组、联合用药各组及顺铂组瘤重均显著低于对照组, 表明高中剂量苦参碱具有较好的抗肿瘤作用, 联合顺铂药物效果更佳。有研究表明苦参碱联合顺铂可显著抑制 H22 荷瘤小鼠肿瘤生长, 并可在某种程度上提高患者生存率, 同本组研究结果类似^[6-10]。

本研究发现顺铂组对 U14 荷瘤小鼠胸腺指数及脾脏指数显著低于对照组, 这与其较明显的抑瘤率形成鲜明对比, 提示顺铂在抑瘤的同时亦对机体免疫器官及功能造成了负面影响,

而苦参碱各剂量组胸腺指数、脾脏指数则显著高于对照组和顺铂组, 表明苦参碱在抑瘤的同时, 可促进小鼠免疫作用的继续发挥, 起到增强荷瘤宿主免疫功能的作用。胸腺及脾脏是哺乳动物重要的免疫器官, 在机体内有肿瘤时, 多种途径作用下免疫功能可出现不同程度的抑制状态, 而苦参碱所具有的抑制肿瘤生长且增强免疫功能的特性给今后肿瘤治疗提供了更多可能^[11-15]。

另外, 本研究中联合高剂量组肌酐、尿素氮水平相比顺铂组显著降低, 而联合中剂量组相比顺铂组则减少, 低剂量组未有明显下降, 表明适度联合苦参碱可协同强化顺铂的抗肿瘤作用, 并可降低顺铂的毒副作用, 有实验证明苦参碱联合 5-FU 对肿瘤抑制作用较优, 且不会损伤静止期骨髓干细胞, 也即降低了 5-FU 对机体所造成的毒副作用, 与本研究有相似之处^[16-20]。

总之, 一定浓度的苦参碱对小鼠宫颈癌 U14 移植瘤有抑

制作用,同顺铂联合应用可增强顺铂的抗肿瘤效果,并减轻后者的毒副作用,对机体免疫力提升亦有促进。该研究为抗肿瘤药物开发及临床肿瘤治疗提供了较好的思路,值得深入探讨。

参考文献(References)

- [1] Luo C, Zhong HJ, Zhu LM, et al. Inhibition of matrine against gastric cancer cell line MNK45 growth and its anti-tumor mechanism[J]. Mol Biol Rep, 2012, 39(5): 5459-5464
- [2] Wang L, You Y, Wang S, et al. Synthesis, characterization and in vitro anti-tumor activities of matrine derivatives [J]. Bioorg Med Chem Lett, 2012, 22(12): 4100-4102
- [3] Qin XG, Hua Z, Shuang W, et al. Effects of matrine on HepG2 cell proliferation and expression of tumor relevant proteins in vitro [J]. Pharm Biol, 2010, 48(3): 275-281
- [4] Liu, TY, Song, Y, Chen, H, et al. Matrine Inhibits Proliferation and Induces Apoptosis of Pancreatic Cancer Cells in Vitro and in Vivo[J]. Biological & pharmaceutical bulletin, 2010, 33(10): 1740-1745
- [5] 魏世杰, 刘梦英, 杨文成, 等. RP-HPLC 法测定犬体内苦参碱血药浓度[J]. 宁夏医科大学学报, 2013, 35(5): 594-596
Wei Shi-jie, Liu Meng-ying, Yang Wen-cheng, et al. RP-HPLC method determined blood drug concentration of Matrine in dogs[J]. Journal of Ningxia Medical University, 2013, 35(5): 594-596
- [6] Ma L, Wen S, Zhan Y, et al. Anticancer effects of the chinese medicine matrine on murine hepatocellular carcinoma cells.[J]. Planta medica, 2008, 74(3): 245-251
- [7] Wang Z, Zhang J, Wang Y, et al. Matrine, a novel autophagy inhibitor, blocks trafficking and the proteolytic activation of lysosomal proteases[J]. Carcinogenesis, 2013, 34(1): 128-138
- [8] Yu HB, Zhang HF, Li DY, et al. Matrine inhibits matrix metalloproteinase-9 expression and invasion of human hepatocellular carcinoma cells[J]. J Asian Nat Prod Res, 2011, 13(3): 242-250
- [9] Chen X, Cao Y, Lv D, et al. Comprehensive two-dimensional HepG2/cell membrane chromatography/monolithic column/ time-of-flight mass spectrometry system for screening anti-tumor components from herbal medicines[J]. Journal of chromatography,A, 2012, 1242: 67-74
- [10] Zhu P, Chen JM, Guo HM, et al. Matrine inhibits disturbed flow-enhanced migration via downregulation of ERK1/2-MLCK signaling vascular smooth muscle cells [J]. Ann Vasc Surg, 2012, 26(2): 268-275
- [11] Liu Z, He D, Zhang X, et al. Neuroprotective effect of early and short-time applying sophoridine in pMCAO rat brain: Down-regulated TRAF6 and up-regulated p-ERK1/2 expression, ameliorated brain infarction and edema[J]. Brain Res Bull, 2012, 88(4): 379-384
- [12] Li LQ, Li XL, Wang L, et al. Matrine inhibits breast cancer growth via miR-21/PTEN/Akt pathway in MCF-7 cells [J]. Cell Physiol Biochem, 2012, 30(3): 631-641
- [13] Zhang J, Li Y, Chen X, et al. Autophagy is involved in anticancer effects of matrine on SGC-7901 human gastric cancer cells. [J]. Oncology reports, 2011, 26(1): 115-124
- [14] Liang CZ, Zhang JK, Shi Z, et al. Matrine induces caspase-dependent apoptosis in human osteosarcoma cells in vitro and in vivo through the upregulation of Bax and Fas/FasL and downregulation of Bcl-2[J]. Cancer Chemother Pharmacol, 2012, 69(2): 317-331
- [15] Fu Q, Fang Q, Feng B, et al. Matrine-imprinted monolithic stationary phase for extraction and purification of matrine from Sophorae flavescentis Ait[J]. J Chromatogr B Analyt Technol Biomed Life Sci, 2011, 879(13-14): 894-900
- [16] Zhang B, Liu ZY, Li YY, et al. Antiinflammatory effects of matrine in LPS-induced acute lung injury in mice [J]. Eur J Pharm Sci, 2011, 44(5): 573-579
- [17] Zhang HF, Shi LJ, Song GY, et al. Protective effects of matrine against progression of high-fructose diet-induced steatohepatitis by enhancing antioxidant and anti-inflammatory defences involving Nrf2 translocation[J]. Food Chem Toxicol, 2013, 55: 70-77
- [18] Li Y, Yuan B, Fu J, et al. Adsorption of alkaloids on ordered mesoporous carbon [J]. J Colloid Interface Sci, 2013, 408: 181-190
- [19] Li Y, Zhang J, Ma H, et al. Protective role of autophagy in matrine induced gastric cancer cell death [J]. Int J Oncol, 2013, 42 (4): 1417-1126
- [20] Peng Y, Guo CS, Li PX, et al. Immune and Anti-oxidant Functions of Ethanol Extracts of Scutellaria baicalensis Georgi in Mice Bearing U14 Cervical Cancers [J]. Asian Pac J Cancer Prev, 2014, 15(10): 4129-4133