doi: 10.13241/j.cnki.pmb.2018.07.011

# 沙漠干热环境创伤失血性休克猪模型的氧代谢特点\*

康 燕1 沈才福234 刘江伟24 是文辉2 夏 亮2

(1 解放军 69240 部队医院 新疆 乌鲁木齐 830000;2 新疆军区总医院新疆特殊环境医学重点实验室 新疆 乌鲁木齐 830000;
 3 解放军 69223 部队 新疆 阿克苏 842300)

摘要目的:探讨沙漠干热环境创伤失血性休克猪的氧代谢特点。方法:选择长白仔猪40头,随机分为四组:常温假手术组(NS)、常 温创伤失血性休克组(NTHS)、干热假手术组(DS)、干热创伤失血性休克组(DTHS),分别置于相应的环境暴露3小时后,进行麻醉, 动静脉置管,NTHS组和DTHS组分别自剖腹术后,行左下叶1/4肝脏切除及脾切除术后,再快速放血至平均动脉压(MAP)降至 45± 5mmHg;NS组和DS组仅行腹中线剖腹术。持续检测计算动脉、混合静脉氧饱和度、氧含量及氧输送(DO<sub>2</sub>)、氧耗(VO<sub>2</sub>)、氧摄 取率(O<sub>2</sub>ER)和动脉血乳酸(Lac)。结果:整个病程中,各组动脉氧饱和度均无显著变化。DTHS组混合静脉氧饱和度和氧含量均较相 同时间点的其他各组低,DO<sub>2</sub>、VO<sub>2</sub>、O<sub>2</sub>ER均显著高于常温环境组(P<0.05)。模型成功后,NTHS组和DTHS组DO<sub>2</sub>均经历"下降-代偿-稳定"的过程,但DTHS组短暂稳定后立即呈进行性快速下降至到动物死亡。在实验过程中,DTHS组各时间点氧摄取率 (O<sub>2</sub>ER)均高于相同时间点的其他组,差异具有统计学意义(P<0.05)。NTHS组和DTHS组氧O<sub>2</sub>ER均在休克后0h出现明显变化, 而动脉血乳酸(Lac)在休克后1.5h才出现明显变化,但DTHS组动脉Lac增高较NTHS组升高更加明显(P<0.05),且进展迅速。结 论:(1)沙漠干热环境创伤失血性休克较高的氧代谢,是机体代偿能力弱、病程变化快的重要原因之一;(2)VO<sub>2</sub>、O<sub>2</sub>ER等直接氧代 谢指标可作为早期评估监测机体氧代谢的敏感指标;(3)血Lac浓度可能是反映干热环境创伤失血性休克严重程度的重要指标。 关键词:沙漠;干热环境;创伤失血性休克;猪;氧代谢

中图分类号: R-33; R605.971 文献标识码: A 文章编号: 1673-6273(2018) 07-1253-06

# The Characteristics of Oxygen Metabolism in Swine Model with Traumatic Hemorrhagic Shock in Dry-heat Environment of Desert\*

KANG Yan', SHEN Cai-fu<sup>23</sup>, LIU Jiang-wer<sup>2</sup>, SHI Wen-hur, XIA Liang<sup>2</sup>

(1 The No.69240 Army Hospital, Urumqi, Xinjiang, 830000, China;

2 The Key Labortary of the Special Environmental Medicine of Xinjiang, The General Hospital of Xinjiang Military Command of the

PLA, Urumqi, Xinjiang, 830000, China; 3 The No.69223 Army, Akesu, Xinjiang, 842300, China)

**ABSTRACT Objective:** To establish investigate the characteristics of oxygen metabolism were compared and analyzed. **Methods:** 40 cases of Landrace piglets were randomly and equally divided into the normal temperature sham operation (NS) group, normal temperature traumatic hemorrhagic shock (NTHS) group, dry-heat environment sham operation (DS) group, dry-heat environment traumatic hemorrhagic shock (DTHS) group. After being exposured to each environment for 3h respectively, the swine were anaesthetized and catheterized, and then laparotomy were performed, after laparotomy, NTHS group and DTHS group underwent left lower lobe 1/4 liver resection and splenectomy, further rapid bleeding to mean arterial pressure (MAP) was approached  $45\pm 5$  mmHg to establish the traumatic hemorrhagic shock model, NS group and DS group underwent laparotomy alone. The arterial, mixed venous oxygen saturation and oxygen content, oxygen delivery (DO<sub>2</sub>), oxygen consumption (VO<sub>2</sub>), oxygen uptake rate (O<sub>2</sub>ER) and arterial blood lactate (Lac) were continuously measured. **Results:** There was no significant change in the arterial oxygen saturation during the whole course of the disease. The mixed venous oxygen saturation and oxygen content of the DTHS group were lower than the other three groups at the same time point. After being exposured for 3 h in dry-heat environment, DO<sub>2</sub> and VO<sub>2</sub> were significantly higher than those in the normal temperature environment group (P<0.05). After the traumatic hemorrhagic shock model were successfully established, the DO<sub>2</sub> of NTHS and DTHS groups underwent the process of "falling-compensatory-stabilization" in each group, but in the DTHS group, after shorter stable stage, the DO<sub>2</sub> was falling quikly to the death. O<sub>2</sub>ER was significantly higher in the DTHS group than other three groups at the same time point (P<0.05). In theNTHS and DTHS group, the O<sub>2</sub>ER had significant changed early in the just establishment of the shock model (0 h). How-

沈才福(1988-),男,硕士,研究方向:特殊环境战创伤研究,E-mail:shencaifu88@163.com

<sup>\*</sup>基金项目:解放军总后勤部面上基金项目(CLZ13J003)

作者简介:康燕(1971-),女,本科,副主任医师,研究方向:超声诊断和军事医学研究,E-mail:ky330164406@163.com

<sup>△</sup> 通讯作者:刘江伟(1970-),男,博士后,主任医师,教授,博士生导师,研究方向:特殊环境战创伤及普外科临床与基础研究,

E-mail: ljw273273@163.com, 电话: 0991-4992552;

<sup>(</sup>收稿日期:2017-08-02 接受日期:2017-08-25)

ever, the level of arterial blood lactate had significant changes until 1.5 h after the establishment of shock model. And the level of lactate was higher in the DTHS group than that of the NTHS group at the same time point. Conclusion: (1) The traumatic hemorrhagic shock in desert dry-heat environment could induce higher oxygen metabolism, which is one important reasons for the weak compensatory ability and rapid deterioration of the course of disease; (2) Direct oxygen metabolism (such as  $VO_{2}$ ,  $O_{2}ER$ ) can be use as a sensitive index for early assessment of oxygen metabolism monitoring; (3) Blood Lac concentration can be used as an important index to assess the serious degree of shock of traumatic hemorrhagic shock in the dry heat environment of desert.

Key words: Dry-heat environment; Trauma; Hemorrhagic shock; Swine; Oxygen metabolism

Chinese Library Classification(CLC): R-33; R605.971 Document code: A Article ID: 1673-6273(2018)07-1253-06

# 前言

失血性休克是由于多种致伤原因导致的血管内血容量急 剧减少,氧输送不足,重要组织器官灌注不足,细胞缺氧,最终 导致多器官功能障碍甚至衰竭的临床常见危急重症印。本课题 组前期的研究显示沙漠干热环境的火器伤及创伤失血性休克 对机体损伤发生早、程度重、生存时间短等特点[24]。研究表明组 织氧代谢障碍是导致创伤失血性休克患者死亡率高的重要原 因<sup>19</sup>,但目前国内外尚未见沙漠干热环境创伤失血性休克的氧 代谢特点相关的文献报道。因此,本研究通过建立沙漠干热环 境和常温环境创伤失血性休克猪模型对比研究沙漠干热环境 创伤失血性休克过程中氧代谢特点,以期为临床病情评价及救 治提供理论依据。

## 材料与方法

#### 1.1 实验动物及主要材料

雄性长白仔猪:购自新疆天康畜牧生物技术股份有限公 司;模拟环境在西北特殊环境人工实验舱(新疆军区总医院研 制)内进行;麻醉呼吸机 EX-60:深圳迈瑞生物医疗电子股份有 限公司; 血气分析仪:GEM Premier 3000,USA; 心电监护仪 (T8): 深圳迈瑞生物医疗电子股份有限公司;BL-420 生物测温 仪:中国成都泰盟;血气针:北京海富达科技有限公司,实验经 新疆军区总医院实验动物伦理委员会审查通过。

## 1.2 实验分组

本地雄性长白仔猪 40 头(25 kg-35 kg,7-9 周),随机分为常 温假手术组(Normal temperature sham group, NS 组, n=10), 常温 创伤失血性休克组 (Normal temperature traumatic hemorrhagic shock group, NTHS 组, n=10), 干热假手术组 (Dry-heat sham group, DS 组, n=10), 干热创伤失血性休克组(Dry-heat traumatic hemorrhagic shock group, DTHS 组, n=10)。

### 1.3 环境设置及麻醉

在人工实验舱内设置环境条件,常温环境:温度 25℃± 1℃,湿度35%±5%;沙漠干热环境;温度40.5℃±0.5℃,湿度 10%± 2%。按以上分组将实验动物分别置于已提前达到相应环 境的实验舱内分别暴露3小时。诱导麻醉使用氯胺酮20 mg/kg、阿托品 0.05 mg/kg 肌肉注射;使用 1.5%-3%的七氟烷混 入 50%的氧气中维持麻醉,维持 BIS 值在 40-60 之间。

#### 1.4 置管及监护

连接心电监护仪,BL-420体温监测,连续监测生命体征及 体温。分离暴露右侧颈外静脉置入漂浮导管以抽取混合静脉血 液样本;右侧股动脉插管监测动脉血压,左侧股动脉用于放血

及血液样本的收集。

## 1.5 模型建立

由于猪具有一个约为人类三倍大小的脾脏,储存较多血 液,创伤失血性休克时可形成自体输血,因此两种环境的创伤 失血性休克猪都将行全脾切除术<sup>66</sup>。沿腹中线开腹、膀胱造瘘。 NTHS 组和 DTHS 组行左下叶 1/4 肝脏及全脾切除术,并输入 三倍脾重的乳酸林格氏液<sup>[7,8]</sup>。休克模型采用固定血压的中度 创伤失血性休克模型<sup>99</sup>,自髂外动脉快速放血致平均动脉压 (MAP)45± 5 mmHg, 稳定 20 min 后记录休克 0 时, 稳定期间可 通过再放血或者静脉输入乳酸林格氏液,稳定目标血压。NS组 和 DS 组按照上述方法完成动脉等监测后,行剖腹术及膀胱造 瘘术后结束手术操作,不进行部分肝脏、脾脏切除和放血。创伤 失血性休克组持续观察监测至动物死亡,NS 组和 DS 组在相 应环境创伤 THS 组最长生存时间上延长约 30 min 后安乐处死。

# 1.6 标本收集、检测及氧代谢计算方法

分别于暴露后、休克成功后(计为休克0时)及休克后每30 分钟自左侧股动脉抽取动脉血;血流动力学平均动脉压 (MAP)、心输出量(CO)等从监护设施连接后即开始连续观察记 录,NTHS 组和 DTHS 组直至动物自然死亡,NS 组至暴露后 11h,DS 组至暴露后 4 小时。动脉血液抽取后置于 4℃保温盒 内 10 分钟内送至我院 ICU 血气分析仪 (GEM Premier 3000, USA)检测动静脉血气、乳酸。实验虽行全程对比研究,但本文 为更为直观清晰的表达沙漠干热环境创伤失血性休克特点,制 图时,仅选取与DTHS组对应时间点。

氧代谢指标及未成年猪的体表面积计算公式如下:  $DO_2 = CO (L/min) \cdot CaO_2 (mlO_2/100ml/blood) \cdot 10/[body-weight]$ (kg); VO<sub>2</sub>=  $(CaO_2-CvO_2) \cdot CO (L/min) \cdot 10/ [body-weight] (kg)$ ;  $O_2 ER = DO_2 / VO_2 = (CaO_2 - CvO_2) / CaO_2; Ca/O_2 (ml/dl) = Hb (g/dl) \cdot$ 1.34(ml/g)+ 0.003(ml/dl • mmHg-1) • PO<sub>2</sub>(mmHg)<sup>[10]</sup>; [body surface area]( $m^2$ )=0.073 · [body-weight]  $2/3^{[7]}$ 

#### 1.7 统计学处理

采用 SPSS 23.0 软件进行统计学处理,符合正态分布的计 量资料以均数±标准差(x±s)表示。采用 Sigmaplot12.5 软件作 图。相同时间点两两比较,方差齐时采用单因素方差分析,并以 LSD 进行多重比较,方差不齐时采用 Kruskal Wallis 检验。P<0. 05为差异具有统计学意义。

#### 2 结果

实验过程中,没有手术或麻醉等意外死亡动物。本研究沙 漠干热环境创伤失血性休克组一般在 3.2-3.5 小时死亡, 常温 环境创伤失血性休克组一般在 10.5-11.5 小时死亡,按实验方案,实验舱于实验动物进仓前达到相应环境设置要求,实验过程中环境稳定,无突然幅波动改变等现象。

四组暴露后体重(BWAE)较暴露前体重(BWBE)均有所减少,但暴露前后差异无显著差异(P>0.05);各组暴露前体温,切除肝脏与体重之比(RL/WT)、呼气末二氧化碳(CPE-TO2)及 BIS 值等基础值无显著差异(P>0.05)。

## 2.1 基础数据

Table 1 Cor	Table 1 Comparison of the basic date between different groups				
		Groups			
Normal temperature			Dry-heat temperature		
THO	NG	THO	,		

表1 各组基础数据比较(x± s)

	THS group	NS group	THS group	NS group
Amount(n)	10	10	10	10
BWBE(kg)	29.61± 2.62	29.96± 2.34	29.67± 2.71	31.9± 2.5
BWAE(kg)	29.1± 2.69	29.38± 2.37	29.08± 2.55	31.2± 2.4
Temperature(°C)	39.03± 0.24	39.12± 0.29	38.99± 0.31	39.08± 0.2
C <sub>PE-T</sub> O <sub>2</sub> (mmHg)	39.7± 3.27	39.5± 3.06	40.4± 3.13	40.6± 3.06
RL/WT(g/kg)	49.53± 8.64	N/A	48.85± 6.99	N/A
BIS	48.9± 5.23	51.2± 4.69	39.7± 5.79	50.2± 4.37

Note: BWBE, body weight before exposure; BWAE, body weight after environmental exposure; CPE-TO2, end tidal carbon dioxide; RL/Wt, resection of liver / body weight; N/A, no hepatectomy.

# 2.2 动脉、混合静脉血氧饱和度和含量

本研究中,通气氧浓度为 50%,病程中 4 组动脉氧饱和度 (SaO<sub>2</sub>)基本处于 100%,各组间无显著差异(P>0.05)。而 DS 组混 合静脉氧饱和度(SvO<sub>2</sub>)始终显著低于 NS 组(P<0.05),但两组均 保持各自的相对稳定状态;DTHS 组和 NTHS 组 SvO<sub>2</sub> 随休克 进展呈下降趋势,但 DTHS 组下降更早更快,整个病程两组均 呈显著差异(P<0.05)。实验期间,NS 组动脉氧含量(CaO<sub>2</sub>)基本 保持与暴露后处于同一水平;DS 组与 NS 组未呈现明显差异 (P>0.05),但由于干热环境血液浓缩,DS 组却呈逐渐上升趋势; 0h后 NTHS 组始终低于 NS 组,除 2h时外,其余均呈现显著 差异(P<0.05),DTHS 组 0h后迅速代偿,虽与 NTHS 组未呈显 著差异,但相同时间点始终高于 NTHS 组。观察期间,DS 组混 合静脉氧含量(CvO<sub>2</sub>)低于 NS 组,DTHS 组和 NTHS 组自 0h 后 呈进行性下降趋势,但 DTHS 组下降更早更快。

表 2 各组动脉、静脉血直接氧代谢指标的比较(x± s) Table 2 Comparison of the blood oxygen metabolism index of artery and vein between different groups

		1	10	5		8 1	
	AE	0 h	1 h	2 h	3 h	8 h	10 h
			Arterial oxygen	saturation(SaO <sub>2</sub> )			
N S	0.99± 0.0	1	1	1	1	1	1
NTHS	1± 0.01	1	1	1	1	1	1
D S	0.99± 0.01	0.99± 0.01	1	1	1		
DTHS	0.991± 0.01	1	1	0.99± 0.01	0.99± 0.01		
		1	Mixed venous oxyg	gen saturation(SvO2	2)		
N S	0.81± 0.04	$0.8\pm 0.04$	$0.8\pm 0.04$	0.79± 0.05	$0.78\pm 0.04$	0.68± 0.27	$0.78 \pm 0.03$
NTHS	0.81± 0.03	$0.64 \pm 0.03^{a}$	$0.64 \pm 0.1^{a}$	$0.57\pm 0.08^{a}$	0.54± 0.11ª	$0.37 \pm 0.18^{a}$	$0.19 \pm 0.15^{a}$
D S	$0.7\pm 0.03^{d}$	$0.71\pm~0.05^{\text{cd}}$	$0.71\pm~0.06^{\text{cd}}$	$0.69 \pm 0.04^{cd}$	$0.69 \pm 0.04^{cd}$		
DTHS	$0.71\pm 0.03^{d}$	0.55± 0.02 <sup>b</sup>	$0.61\pm 0.04$	$0.47 \pm 0.07^{\text{b}}$	0.21± 0.11 <sup>b</sup>		
			Arterial oxyger	n content(CaO <sub>2</sub> )			
N S	12.94± 0.61	12.75± 0.35	13.10± 0.55	12.94± 0.55	12.44± 0.37	12.29± 0.48	12.19± 0.48
NTHS	12.94± 0.51	10.92± 0.65ª	11.47± 0.82 <sup>a</sup>	11.47± 0.81	11.38± 0.51ª	$10.83 \pm 1.30^{a}$	10.99± 1.65 <sup>a</sup>
D S	12.26± 1.02	12.53± 0.91°	12.89± 1.01	13.29± 1.03	13.83± 0.93		
DTHS	12.82± 0.71	10.67± 0.60	12.06± 0.77	12.44± 0.96	12.19± 1.16		
			Mixed venous oxy	gen content(CvO <sub>2</sub> )			
N S	10.44± 0.71	10.16± 0.71	10.54± 0.87	10.2± 0.82	9.72± 0.5	9.43± 0.35	9.45± 0.61
NTHS	10.5± 0.63	6.98± 0.59 <sup>a</sup>	7.34± 1.46 <sup>a</sup>	6.55± 1.22 <sup>a</sup>	6.13± 1.57 <sup>a</sup>	4.01± 2.24 <sup>a</sup>	$2.57 \pm 2.63^{a}$
D S	8.6± 0.99 <sup>d</sup>	8.91± 1.18°	9.18± 1.45°	9.23± 1.2°	9.6± 1.19°		
DTHS	9.05± 0.88	5.83± 0.48 <sup>b</sup>	7.35± 0.69	5.79± 0.97	2.82± 1.13 <sup>b</sup>		

Note: AE, after exposure; NS group, Normal temperature sham group; NTHS group, Normal temperature traumatic hemorrhagic shock group; DS group, Dry-heat sham group; DTHS group, Dry-heat traumatic hemorrhagic shock group. P<0.05 NTHS compared with NS group; P<0.05 DTHS compare with NTHS group; P<0.05 DS compare with DTHS group. dP<0.05 DS group compare with NS group.

#### 2.3 氧输送和氧耗

沙漠干热环境暴露后,氧供(DO<sub>2</sub>)、氧耗(VO<sub>2</sub>)均显著高于常 温组(P<0.05)。随血液的丢失,DO<sub>2</sub>均快速下降后逐渐代偿至某 一稳定水平,但 DTHS 组短暂稳定后呈快速进行性下降至动物 死亡,其再次下降之前高于 NTHS 组。DTHS 组 VO<sub>2</sub> 始终高于 DTHS 组,低于 DS 组。组间相同时间点 DO<sub>2</sub>、VO<sub>2</sub> 的差异见图1。



Fig.1 Trends of oxygen delivery and oxygen consumption

Note: DO<sub>2</sub>, oxygen delivery; VO<sub>2</sub>, oxygen consumption; AE, after exposure; NS group, Normal temperature sham group; NTHS group, Normal temperature traumatic hemorrhagic shock group; DS group, Dry-heat sham group; DTHS group, Dry-heat traumatic hemorrhagic shock group. \*P<0.05 NTHS compared with NS group; \*P<0.05 DTHS compare with NTHS group; \*P<0.05 DS compare with DTHS group. dP<0.05 DS group compare with NS group.

#### 2.4 氧摄取率和动脉血乳酸

环境暴露后,干热环境氧摄取率(VO<sub>2</sub>/DO<sub>2</sub>)显著高于常温 环境(P<0.05),DTHS组氧摄取率在0h时突然升高,整个病程 呈进行性上升,且始终高于其他3组。动脉血乳酸暴露前后无 显著差异,休克后,两休克组血乳酸均呈上升趋势,但 DTHS 组 至 1.5 h 后即呈快速进行性升高,较 NTHS 组升高早且快。组间 相同时间点 VO<sub>2</sub>/DO<sub>2</sub>,Lac 的差异见图 2。



# 图 2 各组氧摄取率及血乳酸变化趋势图



Note: DO<sub>2</sub>, oxygen delivery; VO<sub>2</sub>, oxygen consumption; AE, after exposure; NS group, Normal temperature sham group; NTHS group, Normal temperature traumatic hemorrhagic shock group; DS group, Dry-heat sham group; DTHS group, Dry-heat traumatic hemorrhagic shock group. \*P<0.05 NTHS compared with NS group; \*P<0.05 DTHS compare with NTHS group; \*P<0.05 DS compare with DTHS group. dP<0.05 DS group compare with NS group.

# 3 讨论

生理状况下,机体正常的生理活动有赖于组织氧代谢的供 需动态平衡,即氧输送量≥氧耗量<sup>[9]</sup>。当机体由于某种原因或疾 病导致氧供减少和(或)耗氧量增加,都将打破机体氧供需的动 态平衡,导致组织细胞发生缺血缺氧。根据缺氧的时间及严重 程度,可导致组织、器官损伤,直至衰竭致机体死亡<sup>[11]</sup>。创伤失 血性休克的实质是组织灌注不足引起缺血、缺氧,导致组织器 官损伤,甚至多器官功能衰竭<sup>[12]</sup>,但目前国内外尚未见沙漠干 热环境创伤失血性休克的氧代谢特点报道。因此,本研究探讨

#### 了此环境的氧代谢特点。

氧输送是决定氧供给量的关键因素,主要依赖动脉血流量 及动脉氧含量(CaO<sub>2</sub>)<sup>[5]</sup>。热应激导致心率、心输出量显著增加<sup>[13]</sup>, 从而显著增加动脉血流量。本研究结果显示暴露后的NS组和 DS组 CaO<sub>2</sub>未见显著差异,因此干热环境假手术组(DS)和干热 环境创伤失血性休克组(DTHS)环境暴露后氧供高于常温环境 假手术组(NS)和常温环境创伤失血性休克组(NTHS)。休克模型 建立后,全身总血流量迅速减少,导致NTHS组和DTHS组 DO<sub>2</sub>快速下降,继而导致组织缺氧、灌注不足。组织灌注不足、 缺氧的第一个代偿反应是增加心输出量和增加氧摄取率<sup>[5]</sup>,因 此在 DO<sub>2</sub>下降后即出现代偿性升高;热应激与失血性休克两种 应激因素的协同作用,使 DTHS 组 DO<sub>2</sub> 代偿及维持期间均高 于 NTHS 组,但在 1.5-2 h间开始 DTHS 组 DO<sub>2</sub> 低于 NTHS 组,并呈进行性下降趋势。由此可见,干热环境创伤失血性休克 在代偿期维持较高的氧输送,短暂代偿后的进行性下降是导致 机体有限的氧供快速耗尽的原因,也是沙漠干热环境创伤失血 性休克代偿期短、病程进展快的重要原因之一。

氧耗(VO<sub>2</sub>)反映机体的总代谢情况,代表实时组织吸收的 氧量。VO<sub>2</sub>和DO<sub>2</sub>有着类似的病理过程<sup>114</sup>。与DO<sub>2</sub>相似,干热环 境暴露后,DS组VO<sub>2</sub>显著高于NS组,但VO<sub>2</sub>增加幅度较DO<sub>2</sub> 增加幅度大,约2倍左右,提示干热环境暴露后,机体的耗氧量 大幅提高。Haouzi等指出失血性休克后机体氧耗量高于休克前 机体氧耗量<sup>11516</sup>,而本研究结果显示DTHS组VO<sub>2</sub>代偿虽然始 终高于NTHS组,但未能超过其休克前的氧耗量,提示DTHS 组始终保持较高的氧耗量,但其代偿能力可能较NTHS组低。 因此,我们推测沙漠干热环境创伤失血性休克始终保持较高的 组织耗氧量和较低的代偿能力。

氧摄取率(VO<sub>2</sub>/DO<sub>2</sub>,O<sub>2</sub>ER)是反映组织内呼吸的指标<sup>[17]</sup>。机体代谢增强可通过提高氧摄取率来满足机体对氧的需求,增加 组织呼吸,减少混合静脉氧含量(CvO<sub>2</sub>)。本研究结果显示干热 环境暴露后,DS 组 CvO<sub>2</sub> 显著低于 NS 组,而 O<sub>2</sub>ER 显著高于 NS 组,提示干热暴露后机体组织氧摄取增加。与 NTHS 组相 比,DTHS 组 CvO<sub>2</sub> 较早较快的呈进行性降低,而 O<sub>2</sub>ER 较早较 快的呈进行性升高,提示在干热环境和创伤失血性体克的双重 打击下,机体内呼吸增强增快。正常机体都有一定的氧储存能 力,保证机体在遭受不利因素刺激时,使氧供给尽可能的满足 突如其来的耗氧量增加<sup>[18]</sup>。热暴露期间,机体可能已经耗尽体 内生理氧储存,突然的创伤失血性休克加剧机体氧耗量,因此 这可能是 DTHS 组在 0 h 氧摄取率突然升高的主要原因。

正常机体循环乳酸(Lac)浓度保持产生和消耗的动态平衡, 由于应激或缺血会导致乳酸产生增加,或由于乳酸作为底物的 能量代谢障碍,也将导致乳酸浓度增加,乳酸浓度也被认为是 机体氧债大小重要参考依据<sup>[1920]</sup>,动脉乳酸浓度通常被认为是 视估机体休克严重程度的敏感指标<sup>[52021]</sup>。本研究结果显示动脉 乳酸浓度在 DTHS 组自 0.5 h 开始较 NTHS 组显著增高,但二 者均在 1.5 h 即呈明显升高趋势,且后续呈快速增长趋势,而 NTHS 组虽有升高,但较 DTHS 升高幅度和速度较小且慢。因 此,我们推测沙漠干热环境创伤失血性休克的氧负担及酸中毒 发生时间早,变化速度快,机体损伤程度重。

Kopterides 等发现由于组织产生乳酸与循环中乳酸出现有 一定的时间差,血乳酸比组织乳酸出现的时间较晚、浓度相对 较低<sup>[21-23]</sup>。本研究结果显示 DO<sub>2</sub>、VO<sub>2</sub>、O<sub>2</sub>ER 在环境暴露后即呈 现出显著差异,提示机体在干热环境暴露后机体已呈现氧的高 代谢,而环境暴露前后动脉血乳酸无显著变化,乳酸盐的明显 变化出现在休克后 1.5 h, DO<sub>2</sub>、VO<sub>2</sub>等直接氧代谢指标变化与 血乳酸变化呈现不一致变化的可能机制:(1)此过程增大的氧需 求量由机体的本身氧储存补给<sup>[13]</sup>,尚无缺氧发生,无乳酸生成; (2)机体虽有氧供不能满足氧需,已有无氧代谢至乳酸增多,但 此阶段主要集中于组织内,尚未循环入血,因此在环境暴露与 休克初期并无血乳酸增高的现象。因此,我们推测乳酸盐变化 不是反映沙漠干热环境创伤失血性休克的早期敏感指标,而是 反映其严重程度的有用指标。

综上所述,本研究结果表明:(1)沙漠干热环境创伤失血性 休克加剧机体较高氧代谢,致使机体较快耗尽有限氧含量,是 导致代偿能力降低、病程变化快的重要原因之一;(2)干热环境 VO2、O2ER等直接氧代谢指标于暴露后即与常温环境呈现显著 差异,因此其可作为沙漠干热环境创伤失血性休克早期评估监 测机体氧代谢的敏感指标;(3)与直接氧代谢指标相比,血乳酸 升高变化出现较晚,因此血Lac可作为沙漠干热环境创伤失血 性休克评估监测机体休克严重程度的有用指标。

## 参考文献(References)

- [1] Wang L, Liu F, Yan K, et al. Effects of resuscitation with polymerized porcine hemoglobin (pPolyHb) on hemodynamic stability and oxygen delivery in a rat model of hemorrhagic shock[J]. Artif Cells Nanomed Biotechnol, 2017,45(1): 51-57
- [2] 刘江伟,张永久,李泽信,等.常温和千热环境下腹部肠管火器伤动物 模型的建立[J].创伤外科杂志, 2007, 9(5): 408-410
  Liu Jiang-wei, Zhang Yong-jiu, Li Ze-xin, et al. Establishment of porcine models of fire arm wound of intestine in normal temperature environment and dry heat environment [J]. Trauma Surg, 2007, 9(5): 408-410
- [3] 刘江伟,钱建辉,李瑞,等.沙漠干热环境下创伤失血性休克大鼠模型的建立[J].中国比较医学杂志, 2015, 25(2): 30-33 Liu Jiang-wei, Qian Jian-hui, Li Rui, et al. Establishment of a rat model of traumatic hem orrhagic shock in dry hot desert environment [J]. Chinese Journal of Comparative Medicine, 2015, 25(2): 30-33
- [4] 沈才福,刘江伟,钱若筠,等.沙漠干热环境不同温度对创伤失血性休 克猪生存时间的影响[J].实验动物科学, 2016, 33(4): 48-51 Shen Cai-fu, Liu Jiang-wei, Qian Ruo-jun, et al. The Effect of Survival on Traumatic Hemorrhagic Shock Pigs in Different Temperature of Desert Dry-heat Environment [J]. Laboratory Animal Science, 2016, 33(4): 48-51
- [5] Bursa F, Pleva L. Anaerobic metabolism associated with traumatic hemorrhagic shock monitored by microdialysis of muscle tissue is dependent on the levels of hemoglobin and central venous oxygen saturation: a prospective, observational study [J]. Scand J Trauma Resusc Emerg Med, 2014, 22: 11
- [6] Fry D E. Sepsis, systemic inflammatory response, and multiple organ dysfunction: the mystery continues[J]. Am Surg, 2012, 78(1): 1-8
- [7] Borovniklesjak V, Whitehouse K, Baetiong A, et al. Effects of Intraosseous Erythropoietin during Hemorrhagic Shock in Swine [J]. Plos One, 2014, 9(11): e110908
- [8] Larentzakis A, Toutouzas K G, Papalois A, et al. Porcine model of hemorrhagic shock with microdialysis monitoring [J]. Journal of Surgical Research, 2013, 179(1): e177-e182
- [9] Fulop A, Turoczi Z, Garbaisz D, et al. Experimental models of hemorrhagic shock: a review[J]. Eur Surg Res 2013, 50: 57-70
- [10] Martini W Z, Cortez D S, Dubick M A. Comparisons of normal saline and lactated Ringer's resuscitation on hemodynamics, metabolic responses, and coagulation in pigs after severe hemorrhagic shock [J]. Scand J Trauma Resusc Emerg Med, 2013, 21(1): 86

- [11] McKinley T O, McCarroll T, Gaski G E, et al. Shock volume: A patient-specific index that predicts transfusion requirements and organ dysfunction in multiply injured patients [J]. Shock, 2016, 45 (2): 126-132
- [12] 吴超,顾勤,虞竹溪.中心静脉-动脉二氧化碳分压差在反映感染性 休克患者组织灌注及氧代谢状态的临床研究 [J]. 临床急诊杂志, 2016, (5): 353-357

Wu Chao, Gu Qin, Yu Zhu-xi. The clincal research of central venous-arterial carbon dioxide difference as tissue perfusion and oxygen metabolism in patients with septic shock[J]. Journal of Clinical Emergency, 2016, (5): 353-357

- [13] Crandall C G, Wilson T E. Human cardiovascular responses to passive heat stress[J]. Compr Physiol, 2015, 5(1): 17-43
- [14] Haouzi P. Tissue hypoxia during acute hemorrhage [J]. Crit Care, 2013, 17(2): 423
- [15] Haouzi P, Van de Louw A. Uncoupling mitochondrial activity maintains body [Formula: see text] during hemorrhage-induced O<sub>2</sub> deficit in the anesthetized rat [J]. Respir Physiol Neurobiol, 2013, 186(1): 87-94
- [16] Vincent J L, De Backer D. Oxygen transport-the oxygen delivery controversy[J]. Intensive Care Med, 2004, 30(11): 1990-1996
- [17] McNarry M A, Harrison N K, Withers T, et al. Pulmonary oxygen up-

(上接第1252页)

- [12] Baigude H, Rana TM. Strategies to antagonize miRNA functions in vitro and in vivo[J]. Nanomedicine, 2014, 9(16): 2545-2555
- [13] Takada S, Asahara H. Current strategies for microRNA research[J]. Mod Rheumatol, 2012, 22(5): 645-653
- [14] Matsukura S, Osakabe Y, Sekiguchi A, et al. Overexpression ofmicroRNA-155 suppresses chemokine expression induced by Interleukin-13 in BEAS-2B human bronchial epithelial cells [J]. Allergol Int, 2016, 65(S): S17-S23
- [15] X Liu, L Tian, H Wang, et al. Overexpression of MicroRNA-29a regulates nucleus pulposus cells apoptosis in human intervertebral disc degeneration[J]. Chongqing Medicine, 2016, 45(14): 1893-1895
- [16] Y Wu, Y Zhang, L Wan, et al. Construction of microRNA-21 and PTEN eukaryotic expression and short hairpin RNA expression vectors[J]. Journal of Biomedical Engineering, 2013, 30(2): 359-364
- [17] Chen S, Ni M, Yu B, et al. Construction and identification of a human liver specific microRNA eukaryotic expressionvector [J]. Cell Mol Immunol, 2007, 4(6): 473-477
- [18] Du P, Wang L, Sliz P, et al. A Biogenesis Step Upstream of Microprocessor Controls miR-17~92 Expression [J]. Cell, 2015, 162 (4): 885-899

take and muscle deoxygenation kinetics during heavy intensity cycling exercise in patients with emphysema and idiopathic pulmonary fibrosis[J]. BMC Pulm Med, 2017, 17(1): 26

- [18] Benedik P S, Hamlin S K. The physiologic role of erythrocytes in oxygen delivery and implications for blood storage[J]. Crit Care Nurs Clin North Am, 2014, 26(3): 325-335
- [19] Knoller E, Stenzel T, Broeskamp F, et al. Effects of Hyperoxia and Mild Therapeutic Hypothermia During Resuscitation From Porcine Hemorrhagic Shock[J]. Crit Care Med, 2016, 44(5): e264-e277
- [20] Shoemaker W C, Peitzman A B, Bellamy R, et al. Resuscitation from severe hemorrhage[J]. Crit Care Med, 1996, 24(2 Suppl): S12-S23
- [21] Kopterides P, Theodorakopoulou M, Ilias I, et al. Interrelationship between blood and tissue lactate in a general intensive care unit: a subcutaneous adipose tissue microdialysis study on 162 critically ill patients[J]. J Crit Care, 2012, 27(6): 742-749
- [22] Bursa F, Olos T, Pleva L, et al. Metabolism monitoring with microdialysis in the intensive care [J]. Cas Lek Cesk, 2011, 150 (11): 605-609
- [23] Mallat J, Lemyze M, Meddour M, et al. Ratios of central venous-toarterial carbon dioxide content or tension to arteriovenous oxygen content are better markers of global anaerobic metabolism than lactate in septic shock patients[J]. Ann Intensive Care, 2016, 6(1): 10
- [19] Lopes Ide O, Schliep A, de Carvalho AC. The discriminant power of RNA features for pre-miRNA recognition [J]. BMC Bioinformatics, 2014, 15(1): 124-133
- [20] Sablok G, Pérez-Quintero AL, Hassan M, et al. ArtificialmicroRNAs (amiRNAs) engineering - On how microRNA-based silencing meth- ods have affected current plant silencingresearch [J]. Biochem Biophys Res Commun, 2011, 406(3): 315-319
- [21] Ha M, Kim VN. Regulation of microRNA biogenesis [J]. Nat Rev Mol Cell Biol, 2014, 15(8): 509-524
- [22] TW Ke, HL Hsu, YH Wu, et al.MicroRNA-224 suppresses colorectal cancer cell migration by targeting Cdc42 [J]. Disease Markers, 2017, 2014(1): 617150
- [23] AD Zhou, LT Diao, H Xu, et al. β-Catenin/LEF1 transactivates the microRNA-371-373 cluster that modulates the Wnt/β-catenin-signaling pathway[J]. Oncogene ,2017, 31(24): 2968
- [24] Muz B, de la Puente P, Azab F, et al. The role of hypoxia in cancer progression, angiogenesis, metastasis, and resistance to therapy [J]. Hypoxia, 2015, 3: 83-92
- [25] H Cai, X Liu, J Zheng, et al. Long non-coding RNA taurine upregulated 1 enhances tumor-induced angiogenesis through inhibiting microR-NA-299 in human glioblastoma[J]. Oncogene, 2017, 36(3): 318