

doi: 10.13241/j.cnki.pmb.2018.23.030

## 蓝芩口服液联合阿奇霉素对支原体肺炎患儿免疫功能、血清 $\beta$ 防御素-2、D-二聚体的影响\*

刘开云<sup>1</sup> 刘欢<sup>1</sup> 张融<sup>1</sup> 左丹华<sup>1</sup> 黄富宏<sup>2</sup>

(1 扬州大学附属医院儿科 江苏扬州 225001; 2 扬州大学附属医院药剂科 江苏扬州 225001)

**摘要** 目的:探究蓝芩口服液联合阿奇霉素治疗支原体肺炎患儿的临床疗效及对其免疫功能、血清 $\beta$ 防御素-2、D-二聚体水平的影响。方法:选取2014年1月至2016年6月扬州大学附属医院收治的支原体肺炎患儿80例,根据随机数字法将其分为对照组和治疗组。对照组以阿奇霉素常规治疗,治疗组在阿奇霉素常规治疗基础上服用蓝芩口服液。治疗1周后,评价两组患者的临床症状,检测和比较两组免疫功能及血清 $\beta$ 防御素-2、D-二聚体水平的变化。结果:治疗后,治疗组总有效率为95%,显著高于对照组(80%,P<0.05)。两组患儿治疗后血液中的白细胞计数均显著下降(P<0.05),且治疗组患儿治疗前后白细胞计数下调幅度显著高于对照组(P<0.05)。两组患儿治疗后血清CD<sub>3</sub><sup>+</sup>、CD<sub>4</sub><sup>+</sup>、CD<sub>4</sub><sup>+</sup>/CD<sub>8</sub><sup>+</sup>、NK细胞水平均较治疗前显著升高(P<0.05),且治疗组患儿治疗前后以上指标水平变化幅度均明显高于对照组(P<0.05)。治疗后,治疗组与对照组患儿血清 $\beta$ 防御素-2水平分别上调至(562.86±45.38)pg/mL及(607.37±47.26)pg/mL,血清D-二聚体浓度分别下调至(83.28±10.46)pg/mL及(125.94±14.83)pg/mL,治疗组以上指标的变化均显著高于对照组(P<0.05)。结论:蓝芩口服液联合阿奇霉素治疗支原体肺炎患儿的疗效明显优于阿奇霉素常规治疗,且能显著改善患者的免疫功能,降低患儿血清中D-二聚体的水平、升高 $\beta$ 防御素-2的水平。

**关键词:** 蓝芩口服液; 支原体肺炎; 阿奇霉素; 免疫功能;  $\beta$ 防御素-2; D-二聚体

**中图分类号:** R563.15 **文献标识码:** A **文章编号:** 1673-6273(2018)23-4527-04

## Effects of Lanqin Oral Liquid Combined with Azithromycin on the Immune Function, Serum $\beta$ Defensin-2 and D-Dimer Levels of Children with Mycoplasma Pneumonia\*

LIU Kai-yun<sup>1</sup>, LIU Huan<sup>1</sup>, ZHANG Rong<sup>1</sup>, ZUO Dan-hua<sup>1</sup>, HUANG Fu-hong<sup>2</sup>

(1 Department of Pediatrics, the Affiliated Hospital of Yangzhou University, Jiangsu, Yangzhou, 225001, China;

2 Department of Pharmacy, Affiliated Hospital of Yangzhou University, Yangzhou, Jiangsu, 225001, China)

**ABSTRACT Objective:** To investigate the clinical effect of lanqin oral solution combined with azithromycin on the immune function and serum  $\beta$ -defensin-2 and D-dimer levels of children with mycoplasma pneumonia. **Methods:** From January 2014 to June 2016, eighty cases of children with mycoplasma pneumonia in Yangzhou University Hospital were divided into the control group and the treatment group according to the random number method. The control group was treated with azithromycin and the treatment group was treated with lanling oral liquid combined with azithromycin. After one week of treatment, the clinical effect, immune function, levels of  $\beta$ -defensin-2 and D dimer were detected and compared between two groups. **Results:** After treatment, the total effective rate of treatment group was 95%, which was significantly higher than that of the control group (80%, P<0.05). After treatment, the white blood cell count in the both groups were significantly decreased (P<0.05), and the decline degree of white blood cell count in the treatment group was significantly higher than that in the control group (P<0.05). The levels of CD<sub>3</sub><sup>+</sup>, CD<sub>4</sub><sup>+</sup>, CD<sub>4</sub><sup>+</sup>/CD<sub>8</sub><sup>+</sup> and NK cells in the serum of both groups were higher than those in the control group (P<0.05), and the serum levels of these indicators in the treatment group was higher than that in the control group (P<0.05). The serum  $\beta$ -defensin-2 levels in the treatment group and in the control group were increased to (607.37±47.26) pg/mL and (562.86±45.38) pg/mL, and the serum D-dimer levels were decreased to (83.28±10.46) pg/ml and (125.94±14.83) pg/mL. The reduction degree of treatment group was significantly higher than that of the control group (P<0.05). **Conclusion:** The effect of lanqin oral solution combined with azithromycin on the treatment of children with mycoplasma pneumonia was better than that of azithromycin alone, it could improve the immune function of patients and reduce the serum D-dimer level and increase the serum  $\beta$  defensin-2 level.

**Key words:** Lanqin oral liquid; Mycoplasma pneumonia; Azithromycin; Immune function;  $\beta$  defensin-2; D-dimer

**Chinese Library Classification(CLC):** R563.15 **Document code:** A

**Article ID:** 1673-6273(2018)23-4527-04

\* 基金项目:江苏省自然科学基金项目(BK20130432)

作者简介:刘开云(1971-),女,本科,副主任医师,主要从事临床小儿呼吸系统疾病的诊断与治疗工作,

电话:13801455577, E-mail: 704932537@qq.com

(收稿日期:2018-08-06 接受日期:2018-08-30)

## 前言

支原体肺炎是儿童和青少年在春初及冬天寒冷季节最容易患的肺部炎症疾病<sup>[1]</sup>,为临床呼吸内科常见病,具有发病急、病程长、范围广等特点<sup>[2,3]</sup>,严重影响患儿的健康成长与日常生活。若临床医治不及时,将会严重损害患儿呼吸系统,进而诱使心、脑、胃、肝、肾等各个器官发生病变<sup>[4-6]</sup>,威胁患儿的生命。另外,该病的治疗与住院费用也给家庭带来了严重的经济负担<sup>[7]</sup>。因此,开发能缩短病程、廉价而高效的治疗方法仍然是小儿支原体肺炎研究的热点问题。目前,临幊上用于治疗小儿支原体肺炎的药物主要为大环内酯类抗生素,如阿奇霉素、红霉素等<sup>[9,10]</sup>。研究表明蓝芩口服液对于呼吸道感染疾病具有良好的治疗作用<sup>[11]</sup>。本研究选取2014年1月至2016年6月扬州大学附属医院收治的支原体肺炎患儿80例,观察蓝芩口服液联合阿奇霉素对支原体肺炎患儿临床疗效及对患儿免疫功能、血清中β防御素-2、D-二聚体水平的影响,且将与阿奇霉素常规治疗进行对比,旨在为进一步推广中西药结合治疗支原体肺炎患儿提供参考依据。

## 1 资料与方法

### 1.1 一般资料

选择2014年1月至2016年6月来扬州大学附属医院就诊的支原体肺炎患儿80例,经过临幊医师检查发现各患者均出现不同程度的咳嗽、发热、呼吸音粗等临床症状,X射线全胸片检查发现各患儿肺部纹理不同程度的增多且出现紊乱,两侧或单侧下肺出现斑片状的模糊阴影,符合支原体肺炎的诊断标准<sup>[12]</sup>。纳入标准的患儿均无先天性呼吸系统疾病,无心、脑、肝、肾等重要器官的器质性病变,也无其他感染性疾病。

将纳入标准的80例患儿按照随机数字法分为对照组和治疗组,每组40例,治疗组中,男性21例,女性19例,年龄2~11岁,平均年龄(6.2±2.5)岁,病程1~8天,平均病程(2.9±0.6)天,体温37.2~39.4℃,平均体温(37.7±1.5)℃,单侧感染28例,双侧感染12例;对照组中,男性20例,女性20例,年龄1~12岁,平均年龄(6.4±2.7)岁,病程1~9天,平均病程(2.8±0.7)天,体温37.1~39.5℃,平均(37.8±1.4)℃,单侧感染27例,双侧感染13例。两组患儿的性别、年龄、病程、病情、体温等一般资料比较差异均无统计学意义( $P>0.05$ ),具有临床可比性。本研究中所有患儿家长均签署知情同意书,且本研究经扬州大学附属医

院伦理委员会批准。

### 1.2 治疗方法

对照组40例患儿采用阿奇霉素进行常规治疗<sup>[13]</sup>,将每250mg乳糖酸阿奇霉素充分溶解于250mL5%的葡萄糖溶液中,按照患儿体重换算,以每天10mg/kg的剂量进行静脉滴注给药,治疗7天后观察疗效。实验组40例患儿在采用阿奇霉素进行常规治疗的同时,加用蓝芩口服液进行治疗<sup>[11]</sup>,一次1支,一日3次,治疗7天后观察疗效。

### 1.3 指标观察

两组患儿在治疗前后分别抽取静脉血4mL于5mL无抗凝剂的试管中,取其中2mL全血,利用全自动生化仪,测定其中白细胞计数。其余的2mL室温下静置2h后,置于离心机中,4℃于3000rpm下离心10min,吸取上清液,采用流式细胞仪检测血清中CD<sub>3</sub><sup>+</sup>、CD<sub>4</sub><sup>+</sup>、CD<sub>8</sub><sup>+</sup>、NK细胞水平,采用免疫散射比浊法测定血清中D-二聚体的水平,采用酶联免疫检测法,根据β防御素-2试剂盒(南京建成生物有限公司,南京,中国)说明书中的实验操作,测定其血清浓度。

### 1.4 疗效评价

疗效评价标准分为显效、有效、无效。患儿的咳嗽、发热及呼吸音粗等临床症状消退,X射线全胸片检查肺部正常,且白细胞计数(临幊上正常人血液中白细胞计数的标准为(3.5~9.5)×10<sup>9</sup>/L)及体温恢复到正常范围,则判定为显效;若患儿的临床症状、白细胞计数、体温向正常范围靠近,X射线全胸片检查肺部阴影面积减少,则说明该患儿被有效治疗后病情好转;若患儿的以上症状均无明显改善或加重,则判定为无效。

### 1.5 统计学分析

采用SPSS17.0软件进行统计学分析,计量资料数据以平均值±标准差(Mean±s)表示,多组间比较采用One-way ANOVA检验中的Dunnett检验;计数资料以百分比(%)表示,采用卡方检验,以P<0.05为差异具有统计学意义。

## 2 结果

### 2.1 两组患儿疗效比较结果

治疗组40例患者中,29例显效,9例好转,1例无效,临床总有效率可达95%;对照组40例患者,22例显效,10例好转,8例无效,临床总有效率为80%,明显低于治疗组( $P<0.05$ ),详细结果见表1。

表1 两组治疗效果的比较[例(%)]

Table 1 Comparison of the clinical effect between two groups[n(%)]

Groups	n	Effective	Improve	Invalid	Always effective
Treatment group	40	29(72.5)	9(22.5)	2(5)	38(95)
Control group	40	22(55)	10(25)	8(20)	32(80)
P		0.009	0.152	0.011	0.012

Note: compared with the control group,\* $P<0.05$ .

### 2.2 两组患者治疗前后外周血白细胞计数的比较

治疗前,两组患儿外周血白细胞计数比较差异无统计学意义( $P>0.05$ ),有临床可比性。治疗后,两组患儿外周血白细胞计

数均较治疗前显著减少( $P<0.05$ ),且治疗组患儿治疗前后外周血白细胞计数变化幅度显著高于对照组( $P<0.05$ )。详细数据见表2。

表 2 两组患者治疗前后外周血白细胞计数( $10^9/L$ )的比较Table 2 Comparison of the blood white blood cell count ( $10^9/L$ ) between two groups before and after treatment

Groups	n	Before treatment	After treatment	Difference	#P
Treatment group	40	25.41± 2.79	5.84± 0.74*	19.57± 1.58*	0.001
Control group	40	24.85± 2.66	9.73± 1.03#	15.12± 1.39	0.001
*P		0.318	0.007	0.009	

Note: compared with the control group, \*P&lt;0.05; compared with the same group before treatment, #P&lt;0.05.

2.3 两组患儿治疗前后血清中  $CD_3^+$ 、 $CD_4^+$ 、 $CD_8^+$ 、NK 细胞水平的比较

治疗前,两组患儿血清  $CD_3^+$ 、 $CD_4^+$ 、 $CD_8^+$ 、 $CD_4^+/CD_8^+$ 、NK 细胞水平比较差异无统计学意义(P>0.05);治疗后,两组患儿血清

$CD_3^+$ 、 $CD_4^+$ 、 $CD_4^+/CD_8^+$ 、NK 细胞水平均较治疗前显著升高(P<0.05),且治疗组患儿治疗前后血清  $CD_3^+$ 、 $CD_4^+$ 、 $CD_4^+/CD_8^+$ 、NK 细胞水平变化幅度均明显高于对照组血液中。详细数据见表 3。

表 3 两组患儿治疗前后血清  $CD_3^+$ 、 $CD_4^+$ 、 $CD_8^+$ 、NK 细胞水平的比较Table 3 Comparison of the  $CD_3^+$ ,  $CD_4^+$ ,  $CD_8^+$  and NK cell levels in serum between two groups before and after treatment

Groups	n	Times	$CD_3^+$	$CD_4^+$	$CD_8^+$	$CD_4^+/CD_8^+$	NK
Treatment group	40	Before treatment	57.08± 5.83	33.56± 3.27	27.53± 2.46	1.22± 0.12	8.53± 0.96
		After treatment	70.45± 6.65#	40.62± 3.82#	25.11± 2.31#	1.62± 0.14**	14.26± 1.34**
		P	0.002	0.007	0.031	0.005	0.004
Control group	40	Before treatment	56.69± 5.72	33.48± 3.53	27.46± 2.61	1.22± 0.13	8.48± 1.04
		After treatment	67.42± 6.17#	37.23± 3.92#	26.61± 2.53	1.40± 0.14#	11.25± 1.18#
		P	0.011	0.019	0.063	0.015	0.008

Note: compared with the control group, \*P&lt;0.05; compared with the same group before treatment, #P&lt;0.05.

#### 2.4 两组患者治疗前后血清 $\beta$ -防御素-2 水平比较

治疗前,两组患儿血清  $\beta$ -防御素-2 水平分别为(406.58± 36.41)pg/mL 及(411.26± 38.02)pg/mL,差异无统计学意义(P>0.

05);治疗后,治疗组与对照组患儿血清  $\beta$ -防御素-2 浓度分别上调至(562.86± 45.38)pg/mL 及(607.37± 47.26)pg/mL,治疗组的上调幅度显著高于对照组(P<0.05),详细数据见表 4。

表 4 两组患者治疗前后血清  $\beta$ -防御素-2(pg/mL)含量的比较Table 4 Comparison of the serum levels of  $\beta$ -defensin-2 (pg/mL) between two groups before and after treatment

Groups	n	Before treatment	After treatment	Difference	#P
therapy group	40	406.58± 36.41	607.37± 47.26*	196.11± 20.37*	0.001
Control group	40	411.26± 38.02	562.86± 45.38#	156.28± 18.43	0.003
*P		0.372	0.017	0.006	

Note: compared with the control group, \*P&lt;0.05; compared with the same group before treatment, #P&lt;0.05.

#### 2.5 两组患者治疗前后血清 D- 二聚体水平的比较

治疗前,治疗组与对照组患儿血清 D- 二聚体浓度分别为(271.63± 37.29)pg/mL 及(274.82± 35.41)pg/mL,差异无统计学

意义(P>0.05);治疗后,治疗组与对照组患儿血清  $\beta$ -防御素-2 浓度分别下调至(83.28± 10.46)pg/mL 及(125.94± 14.83)pg/mL,治疗组的下调幅度显著高于对照组(P<0.05)。

表 5 两组患者治疗前后血清 D- 二聚体含量的比较(ng/mL, n=60)

Table 5 Comparison of the serum D-dimer levels between two groups before and after treatment(ng/mL, n=60)

Groups	n	Before treatment	After treatment	Difference	P
therapy group	40	271.63± 37.29	83.28± 10.46*	191.54± 20.62*	0.001
Control group	40	274.82± 35.41	125.94± 14.83#	145.69± 17.58	0.006
P		0.347	0.005	0.008	

Note: compared with the control group, \*P&lt;0.05; compared with the same group before treatment, #P&lt;0.05.

### 3 讨论

支原体肺炎是由支原体经呼吸道进入机体肺部,刺激机体

内的单核细胞、淋巴细胞、巨噬细胞，释放多种细胞因子，引起机体正常生理功能失衡，最终诱发肺部的一系列炎症反应。近年来多项研究表明<sup>[14-16]</sup>支原体肺炎机体内免疫功能异常。机体内的细胞免疫的应答细胞主要为T细胞与NK细胞。T细胞是由骨髓造血干细胞分化，迁移至胸腺发育而成。由于T细胞抗原受体与其表面CD<sub>3</sub>分子的结合物为T细胞活化所必需的物质，血液中CD<sub>3+</sub>的水平可代表成熟T细胞的水平。根据T细胞表面糖蛋白CD分子表达产物的不同，可分为CD<sub>4</sub>和CD<sub>8</sub>，临幊上常用CD4<sup>+</sup>/CD8<sup>+</sup>来反应机体免疫功能的状态，CD4<sup>+</sup>/CD8<sup>+</sup>比值升高说明机体免疫功能增强，反之则说明减弱<sup>[17]</sup>。NK细胞为大颗粒淋巴细胞，可直接溶解破坏肿瘤细胞与病毒感染细胞，是机体的第一道防御屏障，发挥着重要的免疫调节作用，因此机体内NK细胞数量在一定程度上反映了机体细胞免疫功能<sup>[18]</sup>。

$\beta$ 防御素-2是由皮肤粘膜上皮细胞合成和分泌的具有特殊结构的内源性抗菌肽，对皮肤粘膜起着一定的保护作用。 $\beta$ 防御素-2可直接杀伤入侵机体的病原微生物，参与机体的免疫反应，减缓机体的炎症反应并加速创伤的修复<sup>[19,20]</sup>。有研究显示<sup>[21]</sup>肺炎患者血清 $\beta$ 防御素-2的浓度显著低于健康人，而治疗后，其血清浓度恢复正常水平。D-二聚体是由交联纤维蛋白降解而来，可特异性代表纤维蛋白产生和纤溶发生的程度。当机体处于高凝血状态时，血浆D-二聚体水平将明显升高，而其含量的高低可反映血液凝血状态及纤溶活性的强弱<sup>[22,23]</sup>。因此，血浆D-二聚体水平的测定则可用于血栓性疾病的辅助诊断。有报道显示<sup>[24,25]</sup>血浆D-二聚体的水平在凝血功能异常的肺炎患者体内明显升高。

本研究以支原体肺炎患儿为研究对象观察蓝芩口服液联合阿奇霉素对其临床疗效及免疫功能、血清中 $\beta$ 防御素-2、D-二聚体水平的影响，研究结果显示蓝芩口服液联合阿奇霉素治疗的总有效率为95%，显著高于阿奇霉素治疗的80%。两组患儿治疗后外周血白细胞计数均显著下降，且治疗组患儿在治疗前后白细胞计数下调幅度显著高于对照组，说明蓝芩口服液联合阿奇霉素对支原体感染的肺炎具有一定的治疗作用，且效果好于阿奇霉素单独治疗。两组患儿治疗后血清中CD<sub>3</sub><sup>+</sup>、CD<sub>4</sub><sup>+</sup>、CD<sub>4</sub><sup>+</sup>/CD<sub>8</sub><sup>+</sup>、 $\beta$ 防御素-2、NK细胞水平均升高，而蓝芩口服液联合阿奇霉素治疗组患儿治疗前后血清中该指标水平变化幅度均高于对照组，说明蓝芩口服液联合阿奇霉素能改善患儿的免疫功能，效果优于阿奇霉素单独治疗。另外，两组患儿治疗后血清中D-二聚体浓度均下调，且蓝芩口服液联合阿奇霉素治疗组的下调幅度显著高于对照组，说明蓝芩口服液联合阿奇霉素能在一定程度上改善患儿的凝血状态。

综上所述，蓝芩口服液联合阿奇霉素治疗支原体肺炎患儿的疗效明显优于阿奇霉素常规治疗，且能显著改善患者的免疫功能，降低患儿血清中D-二聚体的水平、升高 $\beta$ 防御素-2的水平。由于血浆D-二聚体水平在反映患者凝血状态的灵敏度上有限，因此在后期实验中将进一步测定其他凝血功能相关指标，全面阐述蓝芩口服液联合阿奇霉素对支原体肺炎患儿凝血功能的影响。

#### 参考文献(References)

- [1] Onozuka D, Hashizume M, Hagiwara A. Impact of weather factors on Mycoplasma pneumoniae pneumonia[J]. Thorax, 2009, 64(6): 507
- [2] Saraya T. The History of Mycoplasma pneumoniaePneumonia [J]. Frontiers in Microbiology, 2016, 7(662)
- [3] Koichi I. Clinical Features of Severe or Fatal Mycoplasma pneumoniaePneumonia[J]. Frontiers in Microbiology, 2016, 7
- [4] Smith L G, Cunha B A. Mycoplasma pneumonia and its complications [J]. Infectious Disease Clinics of North America, 2010, 24(1): 57
- [5] McLaughlin M, Simou J, Sill J. Mycoplasma Pneumonia Causing Acute Hypoxemic Respiratory Failure[J]. Chest, 2016, 150(4): 117A-117A
- [6] Carrara C, Abbate M, Sabadini E, et al. Acute Kidney Injury and Hemolytic Anemia Secondary to Mycoplasma pneumoniae Infection [J]. Nephron, 2017
- [7] Chung W S, Hsu W H, Lin C L, et al. Mycoplasma Pneumonia Increases the Risk of Acute Coronary Syndrome: A Nationwide Population-Based Cohort Study[J]. Qjm Monthly Journal of the Association of Physicians, 2015, 108(9): 697
- [8] Sibbel S, Sato R, Hunt A, et al. The clinical and economic burden of pneumonia in patients enrolled in Medicare receiving dialysis: a retrospective, observational cohort study [J]. Bmc Nephrology, 2016, 17(1): 199
- [9] Kolosova N, Geppe N, Dronov I, et al. The efficiency of topical antibacterial therapy for bronchitis in children[J]. European Respiratory Journal, 2016, 48(suppl 60): PA1269
- [10] Blix H S, Vestheim D F, Hjellvik V, et al. Antibiotic prescriptions and cycles of Mycoplasma pneumoniae infections in Norway: can a nationwide prescription register be used for surveillance? [J]. Epidemiology & Infection, 2015, 143(9): 1884-1892
- [11] 赵会娟,张桂玲,任文娟.蓝芩口服液治疗小儿上呼吸道感染疗效观察[J].实用医学杂志,2009,25(19): 3325-3325
- [12] Aizawa Y, Oishi T, Tsukano S, et al. Clinical utility of loop-mediated isothermal amplification for rapid diagnosis of Mycoplasma pneumoniae in children [J]. Journal of Medical Microbiology, 2014, 63(2): 248-251
- [13] 李晓品,李艳红,许凤勤,等.阿奇霉素对肺炎支原体肺炎感染患儿Th1/Th2指标的影响研究 [J]. 中华医院感染学杂志, 2016, 26(16): 3797-3799
- [14] Li X P, Li Y H, Xu F Q, et al. Effect of azithromycin on Th1/Th2 Indexes in children with Mycoplasma pneumonia infection [J]. Chin J Nosocomiol, 2016, 26(16): 3797-3799
- [15] Wang X, Chen X, Tang H, et al. Increased Frequency of Th17 Cells in Children With Mycoplasma pneumoniae Pneumonia[J]. Journal of Clinical Laboratory Analysis, 2016, 30(6): 1214-1219
- [16] Marchioro S B, Maes D, Flahou B, et al. Local and systemic immune responses in pigs intramuscularly injected with an inactivated Mycoplasma hyopneumoniae vaccine[J]. Vaccine, 2013, 31(9): 1305-1311
- [17] Wang Z H, Li X M, Wang Y S, et al. Changes in the Levels of Interleukin-17 Between Atopic and Non-atopic Children with Mycoplasma pneumoniae Pneumonia[J]. Inflammation, 2016, 39(6): 1-5
- [18] 严健,原永明,张舒,等.CD<sub>3</sub><sup>+</sup>、CD<sub>4</sub><sup>+</sup>、CD<sub>8</sub><sup>+</sup>T 淋巴细胞亚群在肿瘤患者外周血中检测的临床意义[J].检验医学, 2013, 28(10): 901-903  
Yan Jian, Yuan Yong-ming, Zhang Shu, et al. Clinical significance of peripheral blood CD<sub>3</sub><sup>+</sup>, CD<sub>4</sub><sup>+</sup> and CD<sub>8</sub><sup>+</sup>T lymphocyte subset determination in patients with tumor [J]. Laboratory medicine, 2013, 28(10): 901-903

(下转第 4578 页)

- Nat Med, 2014, 20(5): 548-554
- [31] Spellman PT, Gray JW. Detecting cancer by monitoring circulating tumor DNA[J]. Nat Med, 2014, 20(5): 474-475
- [32] Shin SY, Ki CS, Kim HJ, et al. Mutant Enrichment with 3'-Modified Oligonucleotides (MEMO)-Quantitative PCR for Detection of NPM1 Mutations in Acute Myeloid Leukemia [J]. J Clin Lab Anal, 2015, 29 (5): 361-365
- [33] Lambirth KC, Whaley AM, Blakley IC, et al. A Comparison of transgenic and wild type soybean seeds: analysis of transcriptome profiles using RNA-Seq[J]. BMC Biotechnol, 2015, 15: 89
- [34] Shahi RB, De Brakeler S, De Greve J, et al. Detection of EGFR-TK Domain-activating Mutations in NSCLC With Generic PCR-based Methods[J]. Diagn Mol Pathol, 2014, 23(3): 163-171
- [35] How-Kit A, Tost J. Pyrosequencing (R)-Based Identification of Low-Frequency Mutations Enriched Through Enhanced-ice-COLD-PCR[J]. Methods Mol Biol, 2015, 1315: 83-101
- [36] Galbiati S, Monguzzi A, Damin F, et al. COLD-PCR and microarray: two independent highly sensitive approaches allowing the identification of fetal paternally inherited mutations in maternal plasma [J]. J Med Genet, 2016, 23(3): 163-171
- [37] Liu P, Liang H, Xue L, et al. Potential clinical significance of plasma-based KRAS mutation analysis using the COLD-PCR/TaqMan((R))-MGB probe genotyping method [J]. Exp Ther Med, 2012, 4 (1): 109-112
- [38] Zuo Z, Jabbar KJ. COLD-PCR: Applications and Advantages [J]. Methods Mol Biol, 2016, 1392: 17-25
- [39] Peng N, Zhao X. Comparison of mutations in lung, colorectal and gastric cancer[J]. Oncol Lett, 2014, 8(2): 561-565
- [40] Soejima K, Yasuda H, Hirano T. Osimertinib for EGFR T790M mutation-positive non-small cell lung cancer [J]. Expert Rev Clin Pharmacol, 2017, 10(1): 31-38
- [41] Thress KS, Brant R, Carr TH, et al. EGFR mutation detection in ctDNA from NSCLC patient plasma: A cross-platform comparison of leading technologies to support the clinical development of AZD9291 [J]. Lung Cancer, 2015, 90(3): 509-515
- [42] Chae YK, Davis AA, Jain S, et al. Concordance of Genomic Alterations by Next-Generation Sequencing in Tumor Tissue versus Circulating Tumor DNA in Breast Cancer[J]. Mol Cancer Ther, 2017
- [43] Olsson E, Winter C, George A, et al. Serial monitoring of circulating tumor DNA in patients with primary breast cancer for detection of occult metastatic disease[J]. EMBO Mol Med, 2015, 7(8): 1034-1047
- [44] Aucamp J, Bronkhorst AJ, Peters DL, et al. Kinetic analysis, size profiling, and bioenergetic association of DNA released by selected cell lines in vitro[J]. Cell Mol Life Sci, 2017, 74(14): 2689-2707
- [45] Aung KL, Donald E, Ellison G, et al. Analytical validation of BRAF mutation testing from circulating free DNA using the amplification refractory mutation testing system [J]. J Mol Diagn, 2014, 16 (3): 343-349
- [46] Spindler KLG, Pallisgaard N, Andersen RF, et al. Circulating Free DNA as Biomarker and Source for Mutation Detection in Metastatic Colorectal Cancer[J]. Plos One, 2015, 10(4): e0108247
- [47] Reid AL, Freeman JB, Millward M, et al. Detection of BRAF-V600E and V600K in melanoma circulating tumour cells by droplet digital PCR[J]. Clinical Biochemistry, 2015, 48(15): 999-1002
- [48] Isobe K, Hata Y, Tochigi N, et al. Usefulness of Nanofluidic Digital PCR Arrays to Quantify T790M Mutation in EGFR-mutant Lung Adenocarcinoma [J]. Cancer Genomics & Proteomics, 2015, 12(1): 31-37
- [49] Lavigne JP, Espinal P, Dunyach-Remy C, et al. Mass spectrometry: a revolution in clinical microbiology? [J]. Clin Chem Lab Med, 2013, 51(2): 257-270
- [50] Bratman SV, Newman AM, Alizadeh AA, et al. Potential clinical utility of ultrasensitive circulating tumor DNA detection with CAPP-Seq[J]. Expert Rev Mol Diagn, 2015, 15(6): 715-719

## (上接第 4530 页)

- [18] Cook K D, Kline H C, Whitmire J K. NK cells inhibit humoral immunity by reducing the abundance of CD4<sup>+</sup> T follicular helper cells during a chronic virus infection [J]. Journal of Leukocyte Biology, 2015, 98(2): 153
- [19] Ghosh S K, Gerken T A, Schneider K M, et al. Quantification of human beta-defensin-2 and -3 in body fluids: application for studies of innate immunity[J]. Clinical Chemistry, 2007, 53(4): 757-765
- [20] Han F, Zhang H, Xia X, et al. Porcine β-defensin 2 attenuates inflammation and mucosal lesions in dextran sodium sulfate-induced colitis[J]. Journal of Immunology, 2015, 194(4): 1882
- [21] Scharf S, Zahlten J, Szymanski K, et al. Streptococcus pneumoniae induces human β-defensin-2 and -3 in human lung epithelium[J]. Experimental Lung Research, 2012, 38(2): 100-110
- [22] Takeuchi D, Inai K, Shinohara T, et al. Blood coagulation abnormalities and the usefulness of D-dimer level for detecting intracardiac thrombosis in adult Fontan patients[J]. International Journal of Cardiology, 2016, 224: 139-144
- [23] Romanelli R G, Cellai A P, Lami D, et al. D-dimer and fibrinolytic activity in patients with decompensated liver cirrhosis[J]. Digestive & Liver Disease, 2015, 47(1): e33-e33
- [24] 郭山春,徐传伟,刘玉芹,等.不同类型肺炎支原体肺炎儿童血浆凝血酶节蛋白和D-二聚体的变化 [J]. 中国当代儿科杂志, 2013, 15 (8): 619-622  
Guo Shan-chun, Xu Chuan-wei, Liu Yu-jin, et al. Changes in plasma levels of thrombomodulin and D-dimer in children with different types of Mycoplasma pneumoniae pneumonia[J]. Chin J Contemp Pediatr, 2013, 15(8): 619-622
- [25] Sulhattin A, Serdal U, Gokten B, et al. The Association between Plasma D-dimer Levels and Community-Acquired Pneumonia [J]. Clinics, 2010, 65(6): 593-597